Downloaded from pharmrev.aspetjournals.org by guest on June 15, 2012

Spet

Perchlorate and the Thyroid Gland

J. WOLFF^a

Laboratory of Biochemical Pharmacology, National Institute of Diabetes and Digestive and Kidney Diseases, National Institutes of Health,
Bethesda, Maryland

I.	Introduction	89
II.	The anion transport system	90
	A. Energetics	90
	B. Anion selectivity	91
III.	Chemistry, metabolism, and distribution of perchlorate	91
	A. Chemistry	
	B. Distribution and metabolism of perchlorate	92
IV.	Transport effects of perchlorate	92
	A. Competition for iodide	
	B. Locus of the perchlorate effect	92
	C. Stimulated efflux?	93
V.	Anomalous antigoitrogenic effect of perchlorate	94
VI.	The clinical use of perchlorate	94
	A. Treatment of thyrotoxicosis	94
	B. Perchlorate toxicity	95
VII.	Amiodarone	96
	A. 5'-Deiodinase effects	
	B. Direct thyroid amiodarone toxicity	97
	C. Amiodarone and iodide excess	97
	D. Amiodarone-induced thyrotoxicosis	97
	E. Amiodarone and hypothyroidism	98
VIII.	The perchlorate discharge test	99
	A. Kinetic considerations	99
	1. k ₁ effects	99
	2. k ₋₁ effects	99
	3. k ₂ effects	99
	4. k ₋₂ effects	99
	5. k ₃ effects	99
	B. The iodide-perchlorate test	100
	C. Examples of iodide excess	
	D. Examples of organification defects	100
	E. Ectopic thyroid tissue	102
IX.	Summary	
X.	Acknowledgments	102
ΧI	References	109

I. Introduction

The resurgence of the clinical use of perchlorate as a means of controlling amiodarone-induced thyroid dysfunction has suggested a need for a more careful examination of the actions of this antithyroid anion. To accomplish this, it is essential to understand the chemical, biochemical, and physiological bases for the action of

^a Address for correspondence: J. Wolff, Building B, Room 2A23, National Institute of Health, Bethesda, MD 20892.

perchlorate in the iodine economy of the thyroid gland. Many of its *late* antithyroid effects leading to hypothyroidism will not be discussed as they differ little from those of the thionamides. On the other hand, the *early* effects in the pathway of iodine metabolism differ markedly from those of the thionamides, and it is these aspects that are the subject of this review. Perchlorate competitively interferes with iodide accumulation in the thyroid; as a result, it blocks hormone production and output like the more familiar thionamide drugs but by a

different mechanism. This property has led to a deeper understanding of thyroid gland function, particularly the iodide transport system and is the basis for its clinical usefulness in treating thyrotoxicosis, particularly when induced by an iodide load. For the same mechanistic reason, perchlorate ion is used to discharge accumulated iodide from the thyroid that has iodide organification defects.

II. The Anion Transport System

To place the functions of perchlorate into proper perspective, a brief summary of the iodide concentrating mechanism is required, for it shares this mechanism with perchlorate. One of the crucial specializations of thyroid tissue is its ability to concentrate iodide selectively from the surroundings where the concentration is very low $(10^{-8} - 10^{-7} \text{ M})$ and where the concentration of chloride ions will be of the order of 0.01 to 0.1 m. The thyroid has thus developed a mechanism that can establish intrathyroidal iodide concentrations up to several hundred-fold those of the external milieu. Such concentrations are presumably required to promote efficient hormone synthesis (Wolff et al., 1959, 1972) because patients lacking the ability to concentrate iodide have goiters and are hypothyroid (reviewed in Wolff, 1983; a dozen or so additional cases have been reported since Alberto et al., 1987; Couch et al., 1985; Léger et al., 1987; Vulsma et al., 1991; Wémeau et al., 1991). This genetic defect can be entirely corrected by providing large doses of iodide that enter the gland by diffusion. The ability to concentrate iodide is shared by some other tissues, such as salivary glands, gastric mucosa, and the choroid plexus (Brown-Grant, 1961), some of which have readily accessible secretions that can be sampled for diagnostic purposes (Wolff, 1983). Nevertheless, it is essentially only in the thyroid gland that the newly concentrated iodide can be further metabolized to form thyroid hormone. This selective transport system can be conveniently divided into two mechanistically distinct properties: those aspects dealing with *energetics* and those dealing with *specificity* for anions i.e., recognition. The latter is more directly pertinent to the perchlorate anion. For recent reviews see Ekholm, 1990; Taurog, 1996; Wolff, 1992; Wollman, 1995.

A. Energetics

Thyroidal iodide transport is "active" not only by electrochemical criteria (Chow et al., 1969; Wolff, 1964; Wright and Diamond, 1977), but also by numerous metabolic criteria: it does not occur in the cold, requires $\rm O_2$ (Surks, 1967), and is a function of the adenosine triphosphate (ATP)^b level (Wolff and Halmi, 1963; Mayaan and Rosenberg, 1968) and generation (largely by oxidative phosphorylation but also by glycolysis; Tyler et al.,

1968). Although such agents as the cardiac glycosides, membrane disruptive compounds such as filipin, inhibitors of oxidative phosphorylation, and glycolysis poisons, will inhibit iodide transport, none of these is specific for thyroid tissue. Hence, these agents are not antithyroid compounds. Extensive reviews of iodide transport energetics are available (Bastomsky, 1974; Carrasco, 1993; Halmi, 1961; Wolff, 1989).

WOLFF

One factor whose importance has gradually emerged is the role of Na⁺ transport in energizing iodide transport. The first clue that Na⁺ flux might energize iodide transport was the demonstration that cardiac glycosides potently inhibited iodide accumulation in thyroid slices (Wolff and Maurey, 1958). Subsequently, it was shown that a sodium ion was required for iodide transport (Alexander and Wolff, 1964; Bagchi and Fawcett, 1973; Iff and Wilbrandt, 1963; Saito et al., 1982). A model was proposed in which sodium and iodide ions were cotransported in a system involving the Na+/K+ adenosine triphosphatase (ATPase) (Bagchi and Fawcett, 1973). In these systems the presence of perchlorate invariably inhibited iodide transport. With the development of thyroid cell lines, and more importantly, thyroid membrane vesicles, it became possible to analyze the details of this relation of sodium and iodide transport. O'Neill et al. (1987) and Nakamura et al. (1988) showed with thyroid plasma membrane vesicles that the Na⁺/I⁻ cotransport was electrogenic, with a thermodynamically downhill transport of ~2 Na⁺ ions driving one iodide against its electrochemical gradient into the cell. The ouabain-sensitive Na⁺/K⁺ ATPase normally mediates this gradient. These vesicle systems show a similar $K_{\rm m}$ for iodide as does intact thyroid, and they are sensitive to perchlorate inhibition. As a result of such studies, the molecule responsible for the transport of iodide (and perchlorate) has been renamed the sodium/iodide symporter. Very recently the symporter molecule has been cloned and expressed into normally noniodide-transporting cells; after that, these cells now show perchlorate-sensitive iodide accumulation (Dai et al., 1996; Vilijn and Carasco, 1989); it has subsequently been cloned from human tissue (Smanik et al., 1996). There is significant similarity to other sodium-dependent transport proteins. The rat protein has 618 amino acids, the human transporter has 643 amino acids, and there is an 84% sequence identity between them. There are 12 putative transmembrane domains but neither the locus for sodium flux, the anion selectivity domain, nor the nature of their interaction has, as yet, been identified. It is to be expected that the same molecule can be identified in the other mammalian iodide concentrating tissue such as salivary gland, gastric mucosa, mammary gland, the choroid plexus, and possibly the iodide-concentrating algae such as Fucus species. At the same time, the symporter would be predicted to be absent or defective in patients with congenital goiter due to iodide transport deficiency (Wolff, 1983).

^b Abbreviations: AIT, Amiodarone-induced thyrotoxicosis; Amiodarone, 2-butyl-3-[3,5-diiodo-4-(β -diethylaminoethoxy)-benzoyl]benzofuran; ATPase, adenosine triphosphatase; TPO, thyroid peroxidase; TSH, thyroid-stimulating hormone.

B. Anion Selectivity

The defining characteristic of iodide transport is its very high specificity for iodide with respect to the much more abundant chloride ion in biological systems. Nevertheless, not only iodide but several related monovalent anions are also selected. The ability to be concentrated by thyroid tissue, or to inhibit iodide transport, permits the construction of the following potency series for monovalent anion-based inhibition of iodide transport in thyroid slices (Wolff, 1964; Wolff and Maurey, 1963): $\text{TcO}_4^- \geq \text{ClO}_4^- > \text{ReO}_4^- > \text{SCN}^- > \text{BF}_4^- > \text{I}^- > \text{NO}_3^- > \text{Br}^- > \text{Cl}^-$. Note that iodide is by no means the most highly selected anion.

Although it was long ago pointed out that this series is also the Hofmeister or lyotropic series that describes the salting-out and salting-in properties by simple ions for many proteins by concentrated solutions of neutral salts (Baumann and Metzger, 1949; Baumann et al., 1956; Collins and Washabaugh, 1985), the molecular basis of this effect is still debated. It is not clear whether this anion sequence, measured at very high concentrations, has any necessary mechanistic relation to what goes on, at low concentrations, in the thyroid. Thus, it is important to determine which solution property of the anions determined this series, e.g. crystal radius, hydrated radius, hydration enthalpy, charge density, etc. Strong base anion exchange resins (usually a large cation with weak field) exhibit a marked preference for ClO₄ (compared, for example, to Cl⁻), thus it seemed likely that selectivity for iodide or perchlorate in the thyroid gland might be based on an ion exchange mechanism using a large cation such as a quaternary amine, e.g. arginine (Wolff, 1989). What does this relationship to anion exchange resins imply? The pioneering studies on the chemical basis of cation selectivity by Eisenman (1961, 1962) made it possible to apply equivalent arguments to anion selectivity (Wright and Diamond, 1977). A simplified view of the assumptions of this theory are: (a) Hydration energy (rather than size) is a prime determining factor for selectivity; (b) electrostatic interactions may or may not be outweighed by the hydration energy—that is, the balance between these two forces determines selectivity. The free energies required to remove water from both the fixed cation and the anion determine the total hydration energy; however, because the cation value remains constant for the leaving and entering anion, the difference between the two anions determines the selectivity. Thus, the largest anion will have the highest affinity for the exchanger because it has the lowest hydration energy. By the same token, the radius of the exchanger site will determine its field strength and, hence, the importance of coulombic interactions. The selectivity sequence depends, therefore, on the relation between changes, from one anion to the next, in the hydration energy and the number of water molecules to be desolvated and to changes in electrostatic energy.

It can be shown that such manipulations (Reichenberg, 1966) readily explain the above selectivity series, and the selectivity ratio is $\gg 1$ when comparing perchlorate to iodide and closer to one (less selective) in membrane vesicles (O'Neill et al., 1987), which may have lost some selectivity during preparation. An instructive example of the relation between anionic hydration and electrostatic forces was calculated by Reichenberg (1966) and was compared with thyroid gland selectivities by Wolff (1989). The anion selectivity of Amberlite IRA400, a strong base (weak field) anion exchanger, is explained neither by the conductances nor the coulombic interactions. On the other hand, inclusion of the hydration enthalpies to calculate the selectivities will predict the series correctly. Not all of the hydration shell need be displaced during binding to the exchanger, and the selectivity sequence is maintained when 0.6 (but not 0.5) of the hydration enthalpy remains in the calculation. The excellent correlation between the ion exchanger and the thyroid selectivity strongly suggests that such an exchange mechanism determines selectivity in thyroid tissue. Although the functional ion size is related to the above forces, one should remember that it may per se influence permeability, as is the case in many cation channels. Further details can be found in (Wolff, 1989; Wolff and Maurey, 1962, 1963).

III. Chemistry, Metabolism, and Distribution of Perchlorate

A. Chemistry

Certain chemical properties of perchlorate are important to its usefulness in biological systems and these will be reviewed here. Salts of perchloric acid (the strongest of all common inorganic acids) have been widely used in analytical chemistry: salts of the smaller cations are very soluble, whereas salts of the larger univalent cations K⁺, Rb⁺, and Cs⁺ are less soluble, and perchloric acid has been used in the analysis of K⁺ or the removal of this cation from biological materials. Nevertheless, KClO₄ is sufficiently soluble for clinical use. The anion is a tetrahedron with the four oxygen atoms at the corners and the chlorine atom at the center. The partial molal ionic volume is 44.5 ml/mol at 25°C compared with 36.7 for iodide. The Cl-O bond length is 1.44 to 1.51 Å, and the O-O distance is 2.43 Å (Kugler, 1969). The ionic radius of the iodide ion is 2.16 Å.

Perchlorate is a good oxidizing agent in reactions of the type:

(in acid)
$$ClO_4^- + 2H^+ + 2e^- = ClO_3^- + H_2O \quad E^o = 1.23 \text{ V}$$

(in base) $ClO_4^- + H_2O + 2e^- = ClO_3^- + 2OH^- \quad E^o = 0.36 \text{ V}$

Perchlorate is explosive at high concentrations in acid, and it is dangerous when used with chloric acid for iodine determinations.

In fact, however, perchlorate is stable under conditions where it is used in biological systems, i.e., dilute

solution, avoidance of elevated temperatures, and neutral pH.

B. Distribution and Metabolism of Perchlorate

Extensive studies on the pharmacology of perchlorate and the other Hofmeister anions have been carried out by Eichler and Hackenthal, particularly with respect to excitable membranes (1962). Anhar et al. (1959) and Lewitus et al. (1962) used labeled perchlorate to study its possible accumulation in the thyroid. Its radioactive properties are: 36 Cl (production 35 Cl (n, γ), half-life 3 \times 10^5 years, β^- 0.71 Mev (98%), electron capture 1.14 Mev (2%)). They found concentration of ${}^{36}\text{ClO}_4^{-}$ in rat and rabbit thyroids but, because of the low specific activity, the thyroid to medium concentration ratios of [36ClO₄-] did not exceed 4 to 5. Peak uptake by the thyroid occurred at 4 to 6 hours (Goldman and Stanbury, 1973) and thyroid-stimulating hormone (TSH) increased the tissue/medium ratio of [³⁶ClO₄⁻]. Chow et al. (1969, 1970), using smaller ${\rm ClO_4}^-$ concentrations, obtained much higher relative thyroid concentrations of perchlorate in rats and guinea pigs such that the thyroid/medium or serum ratios for [ClO₄⁻] approached those for iodide. Moreover, they showed that unlabeled perchlorate or iodide had comparable inhibiting effects on the tissue/medium concentration ratios. On the basis of histological measurements and inulin and sulfate spaces, it was possible to determine the intrathyroidal distribution of ³⁶ClO₄⁻ (Chow et al., 1970). It was found that there was active transport of perchlorate from the exterior into the cell and additional further transport from the cell into the follicular lumen that may be facilitated because it exceeded the ³⁶Cl⁻ distribution considerably. These results are comparable to the distribution of iodide within the thyroid. The ratios of the relative concentrations for ${}^{36}\text{ClO}_4{}^-$ in the different cell compartments at the lowest dose of ${}^{36}\text{ClO}_4^{-}$ that could be used were:

	$^{36}\text{ClO}_4^{-}$	$^{36}\mathrm{Cl}^{-}$
Cell/stroma	6.15	0.14
Lumen/stroma	147.0	0.95
Lumen/cell	24.0	6.8

Perchlorate is eliminated from the body primarily in the urine. Peak blood levels after oral intake are reported to occur at 3 h. The halftimes that have been determined for the rat vary from ~ 8 h (95% in 60 hours) (Eichler and Hackenthal, 1962), < 8 h (Kutzim et al., 1980), to ~ 20 h (Goldman and Stanbury, 1973). Despite the high redox potential (see Section III.A.), perchlorate appears to be excreted virtually unchanged both in the rat (Eichler and Hackenthal, 1962) and in man (Anbar et al., 1959). In the latter experiments, it was shown with doubly labeled perchlorate— 36 Cl 18 O $_4$ —that the oxygen atoms of perchlorate do not undergo significant isotopic exchange in vivo, and only ~ 1 to 3% of the 36 Cl appears as ClO $_3$ — or Cl— in the urine.

IV. Transport Effects of Perchlorate

A. Competition for Iodide

The paradigm for studying perchlorate effects derives from earlier studies using thiocyanate ion. The first hint that anions like thiocyanate or perchlorate might act competitively on iodide transport came from an early study by Astwood (1943) that showed that SCN--induced goiter in rats was prevented by extra iodide administration. Franklin et al. (1944) found that SCNblocked iodide accumulation in the thyroid and Vanderlaan and Vanderlaan (1947) showed that iodide ion accumulated in propylthiouracil-blocked thyroids could be readily discharged by SCN-. A more rigorous demonstration of SCN⁻ competition with iodide was provided in mice by Wollman (1956). However, SCN⁻ was shown to be metabolized in the thyroid and it was moderately toxic. Hence, the discovery by Wyngaarden et al. (1952, 1953), that perchlorate ion produced the same effects but was substantially more potent than SCN-, proved to be very important. These authors first showed that perchlorate markedly inhibited iodide accumulation by the rat thyroid and could discharge accumulated iodide in propylthiouracil-blocked glands. It was later shown that perchlorate effects could also occur in the unblocked thyroid (Ingbar and Freinkel, 1956). The competitive nature of the inhibition (i.e., no inhibition at extrapolated infinite iodide concentration) was established using thyroid slices and analysis by double reciprocal or Dixon plots (Wolff and Maurey, 1962, 1963). Similar results were obtained in simple systems such as isolated thyroid cells and vesicles (O'Neill et al., 1987). When the system is truly competitive, then iodide should inhibit perchlorate accumulation just as the latter inhibits iodide accumulation. Attempts to show this were made with ³⁶ClO₄ and excess iodide inhibited this accumulation (Anbar et al., 1959; Lewitus et al., 1962). Because of the specific activity problem with perchlorate, attempts were made to demonstrate the reciprocal competitiveness with iodide by use of high specific activity anions, TcO₄⁻ and ReO₄⁻, that use the same symporter. These studies showed that the $K_{\rm m}$ constants (anion accumulation) and K_i constants (inhibition of accumulation of one anion by another) were identical within the error of the method (Wolff and Maurey, 1962).

There is little or no effect on organification of iodide by perchlorate (e.g. Alexander, 1959, and many other in vitro studies). What little iodide enters the gland appears to be organified normally (Pitt-Rivers et al., 1958). Perchlorate thus provides a useful agent for the study of iodide transport that is not complicated by effects on later metabolic steps.

B. Locus of the Perchlorate Effect

Because of the complex anatomy of the thyroid follicle, it remains to be established where perchlorate inhibition is exerted. One of the loci is at the inward, active, move-

Downloaded from pharmrev.aspetjournals.org by

ment of iodide across the basolateral membrane., i.e., the symporter (fig. 1). The perchlorate effect on iodide accumulation is seen immediately upon entry of labeled iodide, i.e., at a time when not enough iodide would have accumulated to permit significant effects from alterations in exit rate. This basal location of active transport has been shown in intact thyroid tissue, thyroid cells, monolayers, and in thyroid membrane vesicles (Nilsson et al., 1990, 1994; O'Neill et al., 1987; Wolff, 1964; Wollman, 1995). On the other hand, the thyroid is, at least, a three compartment system, and much of the iodide in the intact thyroid is stored in the follicular lumen; one must, therefore, ask whether perchlorate can also act on cell to lumen transport in the thyroid. The work of Chow et al. (1969, 1970) showed that, as for iodide, perchlorate accumulation occurred in two steps: an electrochemically "active" step across the basolateral membrane and an electrically downhill step across the apical membrane into the lumen; however, because the perchlorate distribution into the lumen was greater than that of chloride, more than simple diffusion appears to be involved. The fact that in polarized thyroid cell monolayers this basolateral step for iodide transport can be stimulated by TSH is in agreement with the above interpretation (Nakamura et al., 1988, 1990; Nilsson et al., 1990, 1994).

Efflux of iodide from the lumen to the circulation requires crossing the same two membranes, as in influx, but in the reverse order. We know from early clinical experience that perchlorate promotes a very rapid and nearly complete discharge of iodide from propylthiouracil-treated glands (discussed below). Several kinetic papers have provided additional evidence for perchlorate-mediated increases in iodide efflux from the thyroid gland (Scranton and Halmi, 1965; Surks, 1967). Roc-

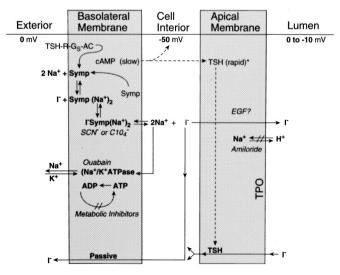


Fig. 1. Scheme of iodide transport in a thyroid follicle (modified from 184). Abbreviations are: Symp = Na/I symporter; r = receptor; Gs = activating G protein; AC = adenylyl cylase; TPO = thyroid peroxidase; EGF = epidermal growth factor. Inhibitors are italicized. Dashed lines indicate direct TSH effects on iodide movement. (*) = rapid efflux across the apical membrane; (**) = rapid efflux from lumen to the exterior.

mans et al. (1977) found that all four transmembrane rates constant had to be adjusted to obtain satisfactory fits in their kinetic simulations. They proposed that perchlorate accomplished this by stimulating counter transport of iodide, but perchlorate does not induce counter transport in reconstituted thyroid membrane vesicles (Saito et al., 1982, 1989). In vitro experiments with thionamide-blocked sheep thyroid slices revealed a two phase efflux of iodide. Both components could be accelerated by perchlorate (fig. 2A). The cellular locus of these two phases is not established. The results are similar to those of sodium ion deprivation (with choline chloride) but are slower when ouabain is used to block the Na⁺/K⁺ ATPase (Salabé and Wolff, unpublished results). This sodium effect had been shown previously (Iff and Wilbrandt, 1963). This discharge should be compared with the classical study on perchlorate discharge in humans (fig. 2B), where the thionamide-blocked thyroid was loaded with radioiodide and measured externally. By current standards, very small doses (10 mg) of perchlorate achieved substantial discharge of thyroidal radioactivity, and 100 mg gave virtually complete discharge (Stanbury and Wyngaarden, 1952). This has become the standard test for organification defects as discussed in Section VIII.

C. Stimulated Efflux?

Can perchlorate-induced iodide discharge from the thyroid lumen be explained by stimulation of efflux, by inhibition of influx alone, or by both? At which of the four transmembrane processes does perchlorate act? Consider the following:

The thyroid cell has a \sim 50 mV negative potential with respect to either the basal or apical compartment (Woodbury and Woodbury, 1963). Thus the interstitium and lumen are roughly isopotential (fig. 1). Reduction of the potential, as is thought to result from TSH stimulation,

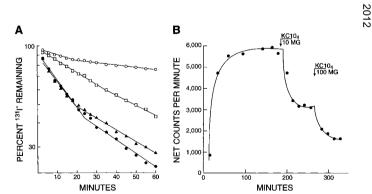


FIG. 2. Perchlorate induced discharge of thyroidal iodide from sheep thyroid slices in vitro and human thyroid gland in vivo. **A.** Experiment of Dr. Helga Salabé (1967). $^{131}\Gamma^-$ prelabeled slices blocked with propylthiouracil were placed into a large volume of medium to minimize recirculation of the released iodide. (\bigcirc) = control; (\bigoplus) = 1 mM ouabain; (\bigoplus) = 1 mM KClO₄; (\bigoplus) = 150 mM choline chloride to replace NaCl; **B.** External counting of a patient's methimazole-blocked (30 mg) thyroid given a tracer dosage of $^{131}\Gamma^-$ at zero time. Arrows indicate oral administration of KClO₄. Reproduced with permission (from Stanbury and Wyngaarden, 1952).

WOLFF

has not been shown to follow exposure to perchlorate but will be important to measure (Williams, 1966).

Iodide movement into the cell from either direction is against an adverse electrical gradient. Across the basolateral membrane there is generally also an adverse concentration gradient; sodium flux and the Na/I symporter overcome this thermodynamic barrier and perchlorate is known to interact here. This will certainly block reentry of external iodide, but is it enough to account for perchlorate discharge of iodide?

Transport out of the cell into the lumen is electrically downhill, but high concentration gradients of lumen/cell iodide (>ten-fold) could, in principle, overcome the favorable potential according to the Nernst relation.

In the reverse direction, movement of iodide from lumen to cell is against the same potential difference, but this may, in contrast to point 3, be overcome by the high concentration gradient for iodide from lumen to cell; it may also be the rate-limiting step for perchlorateinduced iodide efflux. The gradient could be maintained by a very rapid leak of iodide from the cell to the exterior. This is, in turn, maintained by perchlorate-induced inhibition of reentry of iodide from the exterior into the cell, but it remains to be seen whether this is the only effect of perchlorate. Perchlorate added to the apical side of a polarized thyroid cell monolayer is substantially less effective than when added to the basolateral side (Nakamura et al., 1988, 1990). Moreover, perchlorate rapidly increases the secretory response of the thyroid to TSH, including both iodide and iodothyronines (Rousset et al., 1977), and TSH increases iodide efflux before it increases iodide influx (Halmi, 1961); this suggests that additional control points for perchlorate action may exist. We are left then with a readily demonstrated stimulation of iodide efflux from the thyroid produced by perchlorate, not all of which can be readily explained by competitive inhibition of the symporter for influx across the basolateral membrane (Halmi et al., 1960).

V. Anomalous Antigoitrogenic Effect of **Perchlorate**

In experimental animals, perchlorate leads to all the reversible effects expected from the blockade of thyroid hormone synthesis: goiter with histological signs of thyroid stimulation that fails to occur in hypophysectomized animals; thyroidectomy cells in the anterior pituitary; the corresponding changes in thyroid hormone and TSH levels; decrease in tissue QO2, etc. (Enneker et al., 1958; Krüskemper and Kleinsorg, 1954). All these effects are reversed upon cessation of perchlorate administration or with T_3 or T_4 . Nevertheless, perchlorate goiters are not as large as those resulting from propylthiouracil treatment. In the absence of significant organic iodine formation, goiters resulting from the feeding of ClO₄ are about half the size of goiters produced with propylthiouracil. Moreover, the simultaneous feeding of ClO₄ together with propylthiouracil reduces the size of the goiters obtainable with propylthiouracil alone to a size characteristic of the anion (Alexander and Wolff, 1966). These findings have been confirmed in several laboratories (Jolin et al., 1968; Kahl and Bobek, 1971; Matsuzaki and Suzuki, 1981; Ortiz-Caro, et al.,

Efforts to explain this anomalous effect of perchlorate have met with little success:

- (a) It has been known for a long time that large amounts of iodide may reduce but not normalize thyroid size of animals fed thionamide drugs (Alexander and Wolff, 1964; Gassner et al., 1950; Goldberg and Wolff, 1954; Mackenzie, 1947; Milne and Greer, 1962). These changes may be accompanied by modest reductions in the histological signs of thyroid stimulation. It is difficult to see how perchlorate can promote a small iodide leak leading to limited hormone synthesis.
- (b) Propylthiouracil inhibits the peripheral conversion of T4 to T3 but these effects are not produced by the perchlorate. Thus, when the total goitrogenic effect depends on both the intrathyroidal and extrathyroidal effects of drugs, hypothyroidism, and, hence, goitrogenesis, might be less severe with perchlorate despite the fact that the measured TSH levels are the same after both drugs (Alexander and Wolff, 1964; Jolin et al., 1968, 1970; Matsuzaki and Suzuki, 1981). A variant of this explanation stems from the studies of Yamada (1967, 1968), who suggested that perchlorate interfered with the thyroxine plasma protein interaction, thus raising free thyroxine levels and diminishing the goitrogenic stimulus to the pituitary thyrotroph. A direct demonstration of T₄-displacement was not provided.
- (c) Jolin and coworkers (1968, 1970) found that thyroid weight of rats in response to antithyroid agents varies linearly with the circulating insulin level, and that perchlorate treatment is associated with lower insulin levels than is propylthiouracil. Exogenous insulin could restore goitrogenesis to the propylthiouracil level when ClO_4^- was present provided it was administered early enough during goitrogenesis and the adrenal glands are intact (Jolin et al., 1974). An additional factor is the enhancement by perchlorate of TSH sensitivity in the mouse Mackenzie assay (Rousset et al., 1977). Finally, glucose-6-phosphate dehydrogenase levels in thionamide-treated rat thyroids are reduced by perchlorate whereas ornithine decarboxylase is not (Matsuzaki and Suzuki, 1981). It may be concluded, therefore, that perchlorate possesses antigoitrogenic effects in the rat but that the mechanism of this anion effect is complex and remains to be elucidated.

VI. The Clinical Use of Perchlorate

A. Treatment of Thyrotoxicosis

Preliminary clinical studies by Godley and Stanbury (1954) on 24 patients with Graves' disease clearly indicated that such treatment effectively controlled the dis-

ease. Although the use of perchlorate in the treatment of thyrotoxicosis was started in the Unites States, the main early experience with this compound was accumulated in Britain (Crooks and Wayne, 1960; Crooks et al., 1960; Trotter, 1962) and Germany (Krüskemper, 1960a,b, 1962; Wenzel and Lente, 1984). In Germany, iodophobia is so common among some thyroidologists that perchlorate was considered as a possible KI alternative for protection against environmental radioiodine contamination. For various reasons (potential toxicity, short duration of action, different mechanism of action. etc.) this does not seem to be justified. In addition, perchlorate is used in Germany in conjunction with thionamides for prophylaxis against iodine excess derived from coronary angiography (see Fritzsche et al., 1993 for refs.). Such precautions are not generally taken elsewhere. The role of perchlorate in the treatment of thyrotoxicosis can be viewed as having come in two waves. The first enthusiastic period lasted 6 to 7 years and saw well over 1000 (published) patients treated successfully with a side reaction rate not very different from what was seen with thionamides. The appearance between 1961 and 1966 of seven cases of aplastic anemia put a virtual stop to the use of perchlorate, except for single dose-use in perchlorate discharge tests and as an adjunct to pertechnetate scanning. However, with the massive use of amiodarone, a second wave of perchlorate use has been initiated with the lessons learned from earlier reactions kept well in mind.

Initially 400 mg/day taken orally were used in divided (4 to 5/day) doses. The use of frequent daily doses seemed to be important because the blocking effect on iodide uptake was short lived. This led to good, although not very rapid, control of thyrotoxicity with no major toxic reactions save for gastrointestinal disturbances alleviated by dosing with meals (Godley and Stanbury, 1954). It was subsequently found that this dose did not control thyrotoxicity in some patients, and larger doses were initiated (Crooks and Wavne, 1960; Crooks et al., 1960; Trotter 1962). With 600 to 1000 mg KClO₄/day in five divided doses, control was good, generally occurring in 3 to 8 weeks but was attained somewhat more slowly than with methylthiouracil. In some clinics, KClO₄ treatment was started with large initial doses followed by reduction to maintenance levels of 400 to 600 mg/day or to even lower levels (Krüskemper, 1960a.b: Wenzel and Lente, 1984). In a different study using even larger daily doses (1.5 to 2.0 g/day) response was faster (9.4 weeks to remission) than with carbinazole (12.3 weeks), but the number of side reactions was decidedly greater than observed on a regimen of 600 to 1000 mg/day.

One critical difference between thionamides and perchlorate is the danger of using iodides in conjunction with perchlorate, e.g. for the reduction of thyroid vascularity preoperatively. Although this is safe for the thionamides, the fact that iodide and perchlorate are competitive for transport mitigate against its use for the reduction of vascularity much favored by surgeons. The extreme vascularity in patients prepared with perchlorate caused technical difficulties at surgery, hence thionamides, plus iodide is a safer method of preparation for surgery (Cook and Hawe, 1960; Godley and Stanbury, 1954). For the same reason, other iodine-containing medications should be avoided, when possible. Despite the many chemical similarities between the two anions, this is a clear difference from iodide and suggests that the iodide effect on thyroid vascularity may involve the formation of an organic iodine inhibitor not possible with perchlorate (Wolff, 1996).

Perchlorate has been used also when sensitivity to one of the thionamides developed. Combined therapy with perchlorate and thionamide, with the aim of permitting the use of lower concentrations of perchlorate, has been successful (e.g. Blørstad and Vogt, 1962; Røkke and Vogt, 1968); the time to euthyroidism was not appreciably shortened, side reactions are more difficult to assign to a specific drug, and the use of an alternative therapy is lost. Nevertheless, combined therapy has been successful in the occasional cases of iodide-induced thyrotoxicosis after angiography (Fritzsche et al., 1993), in functioning metastatic thyroid carcinoma (Joseph et al., 1994), and particularly in amiodarone-induced thyroid dysfunction (see Section VII.).

B. Perchlorate Toxicity

Experimental studies on perchlorate side reactions in rats at relative dosages much larger than those used in man had shown very little evidence of toxicity (Krüskemper, 1960b), and the clinical findings during the first 6 years revealed primarily minor side effects consisting of nausea or gastritis relieved by administration of the drug with meals, rashes, occasional thyroid enlargement, and leukopenia that was difficult to differentiate from that occurring naturally with thyrotoxicosis. In a review of 881 cases, Krüskemper (1960b, 1962) found these reactions to amount to 3 to 4%, less than seen with the thionamide drugs (see also Røkke and Vogt, 1968). In a more extensive review of the relative toxicity of antithyroid drugs, Trotter (1962) has classified side reactions into three groups: (a) Minor—having an overall incidence of ~10% including drug rashes, drug fever, lymphadenopathy, or nausea. (b) Agranulocytosis with an incidence of $\sim 1\%$; several cases of the nephrotic syndrome were reported at the initial trials but not since. (c) Aplastic anemia with a low incidence. For Trotter (1962), the total incidence of reactions was 2 to 3%, substantially lower than simultaneously estimated reactions to thionamide drugs. As familiarity with perchlorate use increased, dosages were increased in an effort to speed reduction of thyrotoxicity, and it is with these doses (1200 to 2000 mg/day) that there was a large increase in side reaction, going from 2 to 3% to 16 to 18%. Agranulocytosis appeared to be less frequent (0.3% of 1200 cases including single case reports) than in 10,131 cases treated with thionamides (0.94%). The appearance of seven cases of fatal aplastic anemia in 1961 to 1966—four in Britain in 1961 (viz. Trotter, 1962), one in the United States (Krevans et al., 1962), one in Norway (Gjemdal, 1963), and one in Israel (Barzilai and Scheinfeld, 1966)—virtually stopped the use of perchlorate except for single doses used in the perchlorate discharge test (see Section VIII.). It has been popularly assumed that these cases resulted from the use of large dosages of perchlorate, but this is not the case, as all cases followed doses ≤ 1 g/day. On the other hand, most of these patients were treated for periods of 2 to 6 months before aplastic anemia was detected and duration may be a contributing factor. The fact that four of these cases occurred in a cluster raises the question as to whether the perchlorate used at that time may have been contaminated. This question was not followed up at the time. No serious side reactions have been recorded with the resurgent use of perchlorate (Bartalena et al., 1996), and one case with a 22-year maintenance dosage of 200 mg/day had no complications (Connel, 1981). Personal inquiries by Trotter in the 1960s and recently by this author (Germany, Switzerland, Holland and Italy) regarding toxicity after its use for amiodarone-induced thyrotoxicosis at lower dosages and for shorter times. (see Section VII.D.) have not brought to light any additional cases of aplastic anemia or other serious side reactions.

Finally, it must be pointed out that perchlorate readily crosses the placenta (of the guinea pig) and may produce goiter in the fetus, whose thyroids appear to be more sensitive to this anion than that of the mother (Postel, 1957). Crooks et al. (1960) have treated 12 pregnant thyrotoxic women with satisfactory results; 1 neonate had a small, transient goiter.

VII. Amiodarone

The second phase of perchlorate therapy, still ongoing, is based on the special advantages of perchlorate in cases of thyrotoxicosis caused by large iodide loads. Most of these result from the use of the antiarrhythmic agent, amiodarone, but the following arguments are equally applicable to other organic iodine compounds depending only on their half-lives and rates of deiodination. Amiodarone (2-butyl-3-[3,5-diiodo-4-(β-diethylaminoethoxy)-benzoyl]benzofuran) (fig. 3) is an iodinated benzofuran

Amiodarone

Fig. 3. The structure of amiodarone.

derivative containing 37.3% iodine and varying amounts of free iodide.

The drug is stored extensively in many tissues, particularly adipose tissue, and the distribution volume averages 5000 l, leading to a biological half-life of many weeks to 8 months (Holt et al., 1983; Latini et al., 1984, Léger et al., 1984). Its blood level is chronically maintained at 0.7 to 0.9 mg/l (Rao et al., 1986), but values up to 4 mg/l have been reported (Shukla et al., 1991). In one human thyroid, the amiodarone concentrations were about 20 times higher than in blood (Smyrk et al., 1987). In other reports thyroid concentrations varied from 5 to 25 mg/kg, and 75 mg/kg for desethylamiodarone, a metabolite of amiodarone (Haffajee et al., 1985; Holt et al., 1983; Latini et al., 1984). The vast clinical literature, much of it from Europe, where the drug had been more popular than in the United States, reveals that its use is accompanied by numerous side reactions in various organ systems (Mason, 1987). Nevertheless, because the drug is believed to be the only one available for many refractory arrhythmias, these side reactions are often accepted. Among these untoward effects are three of considerable significance for the thyroid. They are: (a) a change in the peripheral metabolism of thyroxine; (b) a direct toxic effect on the thyroid parenchyma: (c) several effects resulting from the excess iodide produced by deiodination of amiodarone. It is important to remember that side effects not easily ascribed to excess iodide alone complicate diagnosis and therapy. We shall summarize these briefly before dealing with the perchlorate-sensitive effects of amiodarone-derived excess iodide.

A. 5'-Deiodinase Effects

A confounding factor in the assessment of the thyroid status in the presence of amiodarone is the fact that amiodarone interferes with the deiodination of T_4 to T_3 by the Type I, propyl-thiouracil-sensitive, 5'-deiodinase (Burger et al., 1976; Franklyn et al., 1985; Melmed et al., 1981; Nadamanee et al., 1989; Singh and Nadamanee, 1983), which accounts for the bulk of peripheral T₃ production. This results in an increase in total and free T_4 , an increase in rT₃, and a corresponding decrease in total and free T₃ in the plasma and most tissues. The Type II, propylthiouracil-resistant 5'-deiodinase of pituitary tissue and brain is apparently little affected by amiodarone. This is probably the most common thyroid abnormality present in amiodarone-treated patients but is generally not accompanied by clinical dysfunction (Newnham et al., 1988). Direct inhibition of the 5'-deiodinase by amiodarone has also been demonstrated in hepatocytes with an IC_{50} of ${\sim}1~\mu\text{M}$ (Aanderud et al., 1984). Such an effect is not produced by an equivalent amount of iodide. Another confounding factor is an increase in the TSH response to thyrotropin-releasing hormone, and a second hormone-binding-related action of amiodarone or its primary metabolic product, desethylamiodarone, is the noncompetitive inhibition of thyroid hormone binding to the β_1 receptor and competitive inhibition to the α_1 receptor, which may cause local hypothyroidism (Bakker et al., 1994; Franklyn et al., 1985; Van Beeren et al., 1995). It has been suggested that some of the salutary effects of amiodarone, particularly in the heart, might be explained by this mechanism.

B. Direct Thyroid Amiodarone Toxicity

A direct toxic effect from intact amiodarone (or its monodeethylated product) has been proposed on the basis of a comparison of postmortem examination of thyroid glands of amiodarone-treated patients who developed thyrotoxicosis; they were found to have thyroid lesions whereas patients without thyrotoxicosis did not (Smyrk et al., 1987). Thyroid damage is seen in needle biopsy material (Capiello et al., 1995) and tissue culture experiments (Brennan et al., 1995). Cell damage suggests a destructive thyroiditis, as shown by decreased chromium retention, loss of tissue contents including thyroglobulin and, hence, thyroid hormones, and decrease in cyclic adenosine 3',5'- monophosphate (cAMP) production (Beddows et al., 1989; Massart et al., 1989). The mechanism of the toxicity is not clearly understood. The lesions produced resemble the damage that is sometimes seen after exposure to excess iodide (Belshaw and Becker, 1973; Many et al., 1992). Reports that both methimazole (Chiovato et al., 1994) and perchlorate (Brennan et al., 1995) prevent thyrocyte damage produced by high concentrations of amiodarone suggest that here too excess iodide may have contributed to the lesions. However, because similar types of damage are also seen in tissues that do not concentrate iodide (Beddows et al., 1989; Chiovato et al., 1994), there can be little doubt that a direct toxic effect is one factor that has to be kept in mind when evaluating amiodarone-induced thyroid dysfunction. Such effects would obviously not be subject to perchlorate competition. As might be expected from the documented tissue damage, several examples of subacute thyroiditis accompanied by thyrotoxicosis, frequently painful, have been encountered (Roti et al., 1992, and references therein). Thyroglobulin levels may be transiently elevated but antithyroid antibodies generally are not. Some of these patients proceed to subclinical hypothyroidism and will show a positive perchlorate or iodide-perchlorate discharge test (see below). Similarly, direct toxic effects may explain the clinical course of patients not responding to perchlorate therapy for amiodarone-induced thyrotoxicosis as well as their response to corticosteroids (Bartalena et al., 1996; Capiello et al., 1995).

C. Amiodarone and Iodide Excess

Amiodarone is metabolized both by the loss of one of the ethyl groups and by deiodination, which is the chief degradation product of interest here. Approximately 10% of the organic iodine is said to be released as iodide per day (Broekhuysen et al., 1969), but it is not known in which organ the bulk of the deiodination occurs. Plasma inorganic iodide concentration was found to rise from $0.05~\mu\mathrm{M}$ to $2.2~\mu\mathrm{M}$, and urinary iodide excretion increased from 0.25 µmol/mmol creatinine to 7 µmol/mmol creatinine or more (Rao et al., 1986). As a result, the absolute thyroid iodine uptake increased to a maximum of 55 nmol/hour. This, in turn, increases the thyroid iodine content as measured by X-ray fluorescence and tends to be greater in patients who become hyperthyroid than in those who become hypothyroid (Fragu et al., 1988: Jonckheer and Deconinck, 1982: Léger et al., 1983. 1984). However, the majority of amiodarone-treated patients have functioning adaptive mechanisms that block or reduce the massive iodide uptakes (Fradkin and Wolff, 1983), and generally only those patients that develop or have developed thyroid dysfunction show these increases. Two forms of dysfunction are observed, roughly as a function of the iodide intake of the population. With adequate dietary iodine supplies, the more common problem is hypothyroidism, whereas in iodinedeficient regions, iodide induced thyrotoxicosis is more common. Put another way, the ratio of hyper/hypothyroidism is <1 in iodine-sufficient areas and >1 in iodinedeficient areas. The incidences reported for thyroid dysfunction vary widely, often because the series are small. from 2 to >30% of amiodarone-treated patients (Harjai and Licato, 1997; Martino et al., 1984; Nadamanee et al., 1989; Trip et al., 1991). An unknown fraction of these patients may have become so because of the direct effects of the drug on the thyroid rather than by iodine excess. Two types of amiodarone effect are important to distinguish (see below) because therapy in cases with damaged thyroid parenchyma will be different. Nevertheless, the success of perchlorate therapy in many of these cases (see below) strongly implicates excess iodide as the cause of the hyperthyroidism.

D. Amiodarone-Induced Thyrotoxicosis

As might be expected, many of the effects of amiodarone resemble those induced by other sources of iodide. Hundreds of cases have been recorded, particularly in Europe and particularly from Martino and coworkers (Fradkin and Wolff, 1983; Martino et al., 1984, 1986a,b, 1987a,b, 1988). Amiodarone-induced thyrotoxicosis (AIT) may appear, often quite suddenly, early or after many years of therapy with amiodarone (Newnham et al., 1988) and may appear many months after withdrawal of amiodarone therapy. It occurs in patients with or without known preexisting thyroid disease. The sex ratio is 3 to 1 males>females; this is in excess of the 2 to 1 ratio of amiodarone-treated patients who remain euthyroid. AIT is not generally accompanied by changes in antibody levels unless preexisting thyroid disease was present (see also Section VII.B.). Because of the large, long half-life, stores of amiodarone, AIT is more difficult to treat than other forms of iodide-induced thyrotoxicosis. Drug withdrawal occasionally is sufficient but gen98 WOLFF

erally remission is slow (Martino et al., 1987a), and some cardiac situations do not permit drug withdrawal. The importance for avoiding thyrotoxicosis in patients with cardiac arrhythmias is self evident, and fatalities from uncontrollable thyrotoxicity have been recorded despite discontinuance of the drug. Occasionally, AIT subsides despite continued therapy. For patients in whom amiodarone cannot be discontinued, examples exist of successful therapy of AIT during continued amiodarone therapy (Reichert and de Rooy, 1989; Trip et al., 1994: Wémeau et al., 1991). The traditional treatment with thionamide drugs such as propylthiouracil or methimazole has had rather limited success. This is perhaps understandable when the following is considered: Assume that thionamide drugs inhibit hormone formation by 95% under normal iodide loads. If now the iodide load is increased by say thirty-fold (which seems guite probable from what has been measured: Rao et al.. 1986), then more organic iodine and thyroid hormone will be synthesized and stored despite what would appear to be adequate therapy for routine Graves' disease. It is for this reason that attempts were made to reduce the excess thyroidal iodide by preventing entry via the symporter and by discharging iodide that might accumulate in the gland from intrathyroidal deiodination.

Martino and coworkers (1986a) compared three treatment modalities in patients with AIT in whom the amiodarone had been discontinued: no treatment, methimazole (40 mg/day) alone, and methimazole plus KClO₄ 1g/day. The unequivocal conclusion was that only the combined drug treatment controlled thyrotoxicity in all cases, and that the time required for attainment of the euthyroid state was greatly reduced (to 1 month or less) compared to those cases that did respond to other treatments. After 40 days, perchlorate was stopped and control was possible with methimazole alone or without further treatment. With combined treatment the return to euthyroidism was a function of the state of the gland before amiodarone: with dysfunction it took 15 to 90 days, without it, it took 6 to 55 days (Martino et al., 1987a). The difficulty in treating thyrotoxicosis in the presence of an iodide load has been noted previously, as has the success of treating Graves' disease rapidly with combined thionamide and perchlorate (Léger et al., 1984; Røkke and Vogt, 1986). It is of interest that the urinary iodine output in the patients in whom amiodarone had been withdrawn showed a brisk increase in the first week or two after institution of the combined therapy. Perchlorate in the absence of amiodarone will also do this (De Groot and Bühler, 1971). These results have been confirmed (e.g. Newnham et al., 1988). In some series the response to combined therapy is slower than above, and there are occasional patients that are refractory, possibly due to direct toxic amiodarone effects on the thyroid parenchyma (see above). Perchlorate dosages have varied between 600 and 1000 mg/day but not more because of the fear of serious side reactions. We are not aware that a minimum effective dosage has been established when used in combination with thionamides.

An unknown fraction of these patients may have become thyrotoxic because of the direct effects of the drug on the thyroid rather than by iodine excess. The latter may not respond to perchlorate and are treated with corticosteroid as for subacute thyroiditis. The distinction between these two types of AIT is now being made and has been classified in table 1 (Bartalena et al., 1996).

E. Amiodarone and Hypothyroidism

Inhibition of thyroid function by excess iodide leading to hypothyroidism and/or iodide goiter has been studied extensively in the past (reviewed in Wolff, 1969). It appeared to be more common in areas of sufficient dietary iodine intake and so it is with amiodarone (Harjai and Licato. 1997: Hawthorne et al., 1985: Martino et al., 1987a,b). Because hypothyroidism is more common in patients with preexisting thyroid dysfunction, many thyroidologists assume that iodide-induced hypothyroidism is presumptive evidence for the unmasking of a preexisting thyroid disorder such as preclinical Hashimoto's thyroiditis. Although this may be the case after amiodarone, hypothyroidism can occur in apparently normal thyroid glands as best as can be established. Of course, when amiodarone induces destructive thyroiditis, it is likely that hypothyroidism may eventually occur (see above). Hypothyroidism after amiodarone can be so severe as to lead to myxedema coma (Mazonson et al., 1984), or it may be merely a laboratory diagnosis (Roti et al., 1992, 1994). An interesting finding is that there may be uninhibited or even increased radioiodine uptake (Martino et al., 1988; Wiersinga et al., 1986). Such patients frequently show a positive perchlorate discharge test indicating incomplete organification of iodide.

The obvious therapy is exogenous thyroid hormone and this has been widely used. Care has to be taken that the amiodarone-induced inhibition of the conversion of T_4 to T_3 is taken into account. It is, of course, especially important to avoid overdosing with T_4 or T_3 in patients who already have arrhythmias. Of greater interest for the present review is the counter-intuitive finding that perchlorate can discharge enough iodide and/or block enough iodide entry into the gland to restore clinical and biochemical euthyroidism in many patients in whom hypothyroidism was induced by amiodarone. This type

	AIT-I	AIT-II
Underlying abnormality	Yes or no	No
Radioiodine uptake	Normal/increased	Decreased
Pathogenesis	Excess iodide	Destructive
Serum IL-6	Slightly increased	Markedly increased
Therapy	Perchlorate (+MMI)	Corticosteroids

of effect was first noted many years ago by Raben (1949) who observed that the inhibition of organic iodine formation caused by excess iodide could be reversed by thiocyanate. In six patients with amiodarone-induced hypothyroidism, 1 g of potassium perchlorate for 9 to 34 days restored euthyroidism with decreased TSH levels and increased FT_4I (Martino et al., 1986b), and was maintained for some time. In others it reverted in 5, 47, and 73 days after cessation of perchlorate therapy but could again be normalized with repeat perchlorate therapy. Similar results have been obtained by others (Van Dam et al., 1993; Wémeau et al., 1991).

How can this paradoxical effect of perchlorate be explained? Excess iodide inhibits many parameters of thyroid function from iodide transport, to cAMP generation, to hormone synthesis, etc. (reviewed in Wolff et al., 1989). In all cases, these effects occur only when oxidation of the accumulated thyroid iodide is permitted. This has led to the postulate that an iodinated organic inhibitor is formed. Recently, several iodinated lipids, iodolactones derived from arachidonic acid, and 2-iodohexadecanal have been identified in the laboratory of Boeynaems in Brussels (viz. Wolff, 1996) as potential inhibitor candidates.

One obvious problem with this proposal would be that the reduction of iodinating species might decrease both protein and lipid iodination equally. The important discovery that lipid iodination required ~10 times more iodide than protein iodination (Pereira et al., 1990) offers a possible explanation for the salutary effect of perchlorate in amiodarone induced hypothyroidism: perchlorate will disproportionately affect lipid iodination while still permitting protein iodination. This is still hypothetical but offers a pathway for thinking about the apparently paradoxical effect of perchlorate.

VIII. The Perchlorate Discharge Test

A. Kinetic Considerations

The significance of the perchlorate discharge test is best understood by considering the following simplified scheme for the early steps of the iodine pathway in the thyroid. In an unblocked gland, the simplest mode for depicting organic iodine formation is:

$$I_{o_{ut}}^{-} \underbrace{\frac{k_1}{k_{-1}}}_{I_{in}} \underbrace{\frac{k_2}{k_{-2}}}_{\text{oxidized}} \text{ (oxidized)} \xrightarrow{\qquad } \text{(organic iodine)}$$

where k_1 and k_{-1} are the influx and efflux rate constants, k_2 is a lump constant for formation of reactive iodine (I° or I⁺), and k_3 is a lump constant for iodination of tyrosine. When iodination proceeds normally $k_2k_3k_{-1} > k_1$ and the intrathyroidal iodide pool is small. When for various reasons $k_1 > k_{-1}k_2k_3$, iodide may accumulate in the thyroid and such a pool is subject to discharge by perchlorate. There are many ways of upsetting the normal steady-state, and this often leads to positive per-

chlorate tests. These are summarized below according to the rate constant involved.

- 1. k_1 Effects. When the thyroid is stimulated but k_1 is increased more than k2 or k3, as may happen in Graves' disease (see below), then dischargeable intrathyroidal iodide may accumulate. Reduction of thyroid mass by surgical or radioiodine ablation may also lead to a buildup of the iodide pool. This may be the result of TSH stimulation of the remaining tissue and the consequent increase in iodide transport as was shown long ago for induction of iodide inhibition by low iodine diets in the rat thyroid (Wolff, 1969). Because the total entry of iodide into the thyroid is a function of [I⁻]k₁, increased iodide loads [I] i.e., increased serum iodide levels, may increase iodide accumulation in the thyroid with or without increases in k₁ This will eventually be in excess of what can be handled by the subsequent iodination steps. This occurs with iodine-rich drugs or diet (seaweed) and does not necessarily imply an iodide effect on the oxidizing system. The relative iodide load effect is also the basis of the iodide-perchlorate test that has greater sensitivity than perchlorate alone for the detection of organification defects (see Section VIII.B.).
- 2. k_{-1} Effects. We are not aware of any examples of reduced iodide efflux leading to iodide accumulation in the gland, but such an effect cannot, in principle, be ruled out even though the main effect of perchlorate is on iodide influx (decreasing k_1).
- $3.\ k_2$ Effects. Oxidation of iodide to reactive forms such as I° or I⁺ requires, as a minimum, thyroid peroxidase (TPO) and a peroxide-generating system. Absent or defective peroxidase (mutations, mislocation, absence of protoporphyrin IX, etc) must lead to the accumulation of dischargeable iodide in the gland as seen, for example, in certain congenital goiters. The classic example of such a block is the attack on the oxidizing system by thionamide drugs. This is, in fact, the finding that started the discharge tests, first with thiocyanate (Stanley, 1949; Stanley and Astwood, 1948) and then with perchlorate (Stanbury and Wyngaarden, 1952). Absence of peroxide could also lead to iodide accumulation. We are aware of only one example of defective peroxide-generating systems (Kasukabe, 1975).
- 4. k_{.2} Effects. In theory, reduction of the oxidized iodine species to iodide, by excess intrinsic thiol groups in the thyroid, for example, might contribute to an enlarged iodide pool. This would resemble the end result of thionamide exposure. We know of no data that has actually demonstrated this. In this regard it is of note that excess iodide leads to an increase in thyroid glutathione (Allen, 1993).
- $5.\ k_3$ Effects. Oxidized iodine requires a protein acceptor (usually thyroglobulin). Some examples of defects in thyroglobulin have been reported and blamed for positive discharge tests, although the reason for the inability to iodinate at least some of the remaining nonhormonogenic tyrosyl residues has remained obscure. Decreased

substrate (thyroglobulin) concentrations also seem unlikely because tyrosyl residues of other proteins in the thyroid could serve as alternate acceptors.

The perchlorate discharge test has been carried out in many variations. Tracer ¹³¹I or ¹²³I is given 1 to 2 hours before perchlorate, neck counts are taken (preferably corrected for neck background), and then 200 to 1000 mg of KClO₄ are given orally. It is not clear that the larger dosages are necessary. At early intervals thereafter (15 to 60 minutes, occasionally 120), neck counts are repeated. Timing is important and should be short to minimize counts from organic iodine in partial defects. Discharge of isotope from the neck is usually rapid in positive cases and may or may not be followed as a function of time to see how low the residual neck count will get. When discharge is slow, e.g., significant only after >2 hours, consideration of additional factors should be entertained. The criteria for positivity vary widely in different clinics and range from a loss of >10% to >30% of the radioactivity accumulated at the time of perchlorate administration. This is, in part, a function of the precise timing and the absolute radioiodine uptake (dietary iodine) of the region (Martino et al., 1984; Trip et al., 1991). The intravenous perchlorate test can be carried out at much shorter time intervals, e.g., 10 minutes after radioiodine and is felt to be significant with 1 to 2% discharge (Gray et al., 1973; Hilditch et al., 1982). It is uncertain whether much is gained by this variation.

B. The Iodide-Perchlorate Test

As detailed in the previous section, administration of an iodide load can increase the intrathyroid iodide pool such that the capacity of the iodinating system may be exceeded more easily. This would be expected to expose marginally sufficient oxidizing systems and lead to a larger fraction of positive perchlorate discharge tests but may become meaningless when the iodide load is so large as to exceed the normal iodinating capacity or provoke its own antithyroid effects. Thus, normal patients may show positive tests when the loading iodide dose reaches ≥ 1 mg (Childs et al., 1950; Stewart and Murray, 1967). In this test, iodide is given along with the radioactive tracer in doses varying from 200 to 1000 µg. The remainder of the test is carried out as above. The large variation in the incidence of positive discharge tests in Hashimoto's thyroiditis (Morgans and Trotter, 1957; Murray and McGirr, 1960; Volpé et al., 1965) led Takeuchi et al. (1970) to compare the standard discharge test with one in which 500 µg of I had been given with the ¹³¹I. Under these conditions normal subjects still showed no discharge, whereas in Hashimoto's thyroiditis (8 patients) the number of positive tests increased from 2 to 8. Other thyroid disorders show similarly increased discrimination for marginal defects (Friis, 1987).

C. Examples of Iodide Excess

Iodide given in excess, either as inorganic iodide or derived from organic precursors (contrast media, amiodarone, etc.) given over prolonged periods of time, leads to iodide goiter, among other problems (Wolff, 1969). Many of these patients exhibit positive perchlorate discharge tests. Although occasional thyroid damage has been demonstrated after iodide, the fact that most of these cases are fully reversible suggests that this must be a minor complication in the majority of such patients. It is not always clear when this is due simply to a load effect, [I⁻]k₁, or indicates inhibition or various thyroid functions (Wolff, 1989). Recovery after cessation of iodide excess suggests a load effect. For amiodarone treatment, simple reflection suggests that hypothyroidism would be more likely to yield positive discharge tests than hyperthyroidism. This has proved to be the case (Martino et al., 1988; Roti et al., 1994).

Of considerable interest is the finding that patients with Graves' disease may have dischargeable iodide pools in their thyroids (Roti et al., 1994). Although most metabolic processes are likely to be accelerated in such glands, this does not occur to the same extent in all of them and the positive perchlorate test suggests, but does not prove, that there is a proportionately greater increase in iodide transport than in organification. The possibility that this may be due to tissue damage (as with coexisting thyroiditis) should be kept in mind.

D. Examples of Organification Defects

Historically, discharge of accumulated intrathyroidal iodide was first demonstrated in thionamide-blocked glands by Stanley and Astwood (1948), who used thiocyanate to compete for iodide. When the advantages of perchlorate became apparent, this anion was substituted and proved extremely effective (as shown in fig. 2A). This figure shows nearly total discharge of label after administration of 100 mg KClO₄, and substantial discharge by as little as 10 mg, leading to the conclusion that the thionamide block was virtually complete. This has been confirmed many times since. In some cases of Graves' disease treated for prolonged periods, a positive iodide-perchlorate discharge test persists 40 days after drug withdrawal (Roti et al., 1994). It is debated whether this is a drug-induced long-term effect or persistence of the hyperthyroid state. Patients treated with lithium salts for bipolar disease often exhibit antithyroid effects and may show a positive iodide-perchlorate discharge test (Andersen, 1973). The locus of lithium action in the thyroid is not established. See table 2 for these and other abnormal discharge tests.

Failure of the oxidizing system of the thyroid may result from a variety of causes (Furth et al., 1967; reviewed in Lever et al., 1983; Medeiros-Neto and Stanbury, 1994). Functional thyroid peroxidase may be deficient or completely lacking as seen in congenital goiters

Downloaded from pharmrev.aspetjournals.org by guest on June

TABLE 2 Positive perchlorate discharge tests

Conditions	Comments	References
Antithyroid compounds		
Thionamides	Test of treatment adequacy	n, ah, ai, aj, ak, al, ao,
Aromatics		h, z,
Lithium	Iodide-perchlorate test only	b,
Iodide excess	Increased load	
Inorganic	Single or prolonged dosing, occasional thyroid damage	l, t, x, ai, aq,
Contrast media	Highly variable $t_{\frac{1}{2}}$	m, x, aq,
Amiodarone	With thyroiditis, thyrotoxicosis, hypothyroidism	e, p, w, aa, ab, ac, ah, ap,
Seaweed	Unsuspected reversible hypothyroidism \pm iodide goiter	s, ag, aq,
Congenital defects		
Congenital goiter	Various defects	c, i, q, ad,
Pendred's syndrome	Peroxidase normal	c, d, k, l, y, ad, af, am,
Thyroiditis		
Hashimoto's		r, ae,
Post-partum	Especially if preceded by hyperthyroid phase	g,
Transport excess $(k_1>k_2)$	Stimulated glands	
Some Graves' disease		
Reduced thyroid mass	Surgical or radioiodine	f, u,
Nontoxic and nodular goiter	Some cases	o, r, v,
Some newborns	Transient	aq,
Ectopic thyroid tissue		
Lingual thyroid	Scintigraphy ¹³¹ I-, ^{99m} TcO ₄ -	a, i, j,
Functioning metastasis	Scintigraphy	t, an,

References: a, Alberto et al., 1987; b, Andersen, 1973; c, Baschieri et al., 1963; d, Bax, 1966; e, Capiello et al., 1995; f, Clark et al., 1990; g, Creagh et al., 1994; h, Edwards et al., 1954; i, El-Desouki et al., 1995; j, Ferrini and Biassoni, 1966; k, Fraser et al., 1960; l, Friis, 1987; m, Fritzsche et al., 1993; n, Godley and Stanbury, 1954; o, Gray et al., 1973; p, Hawthorne et al., 1985; q, Hilditch et al., 1985; r, Hilditch et al., 1982; s, Inada et al., 1983; t, Joseph et al., 1994; u, Kieffer et al., 1965; v, Koutras et al., 1978; w, Lambert et al., 1990; x, Leger et al., 1984; y, Lever et al., 1983; z, Mackenzie, 1947; aa, Martino et al., 1987b; ab, Martino et al., 1986a; ac, Martino et al., 1988; ad, Medeiros-Neto and Stanbury, 1994; ae, Morgans and Trotter, 1957; af, Morgans and Trotter, 1958; ag, Okamura et al., 1994; ah, Roti et al., 1992; ai, Stanbury and Wyngaarden, 1952; aj, Stanley and Astwood, 1948; ak, Stewart and Murray, 1966; al, Takeuchi et al., 1970; am, Trotter, 1960; an, Valenta, 1966; ao, Vanderlaan and Vanderlaan, 1947; ap, Wemeau et al., 1991; aq, Wolff, 1969.

or defective because of its inability to bind one of its substrates. There may be a defect in protoporphyrin IX binding; in certain porphyrinopathies from lead poisoning or hepatic porphyria (Haust and Meagy, 1980), there may be positive discharge tests, but these tend to show a slow release of radioiodine, and other factors may also play a role (Harjai and Licato, 1997). Thyroid peroxidase is normally bound to the apical cell membrane, and mislocation has been proposed as a possible cause of defective organification. In general, however, the structural defects in the peroxidase remain to be elucidated. Finally, a lack of thyroglobulin may be accompanied by normal or high levels of peroxidase and thereby lead to iodide accumulation. Abnormal thyroglobulin has also been implicated, but, as mentioned above, it is difficult to comprehend how all of the solvent-exposed tyrosyl residues of thyroglobulin could have become unavailable for iodination (Floyd et al., 1960). It is of considerable interest that neonatal hypothyroidism associated with a positive discharge test may be transient (Nose et al., 1985). This was ascribed to immaturity of the organification mechanism.

Pendred's syndrome, congenital nerve deafness with goiter and a positive perchlorate discharge test (Fraser et al., 1960), is an example of a partial organification defect in which patients are usually euthyroid. One hundred years after its first description and after a study of nearly 1000 cases, the basis for the goiter and positive discharge test remains obscure. Despite a few reports of defective thyroid peroxidase, most investigators feel that the defect resides elsewhere (Medeiros-Neto and

Stanbury, 1994). It has also been suggested that abnormal thyroglobulin may be associated with the syndrome, but again this is far from established. It is of interest that the presence of a positive perchlorate discharge test has become a *sine qua non* for the diagnosis.

The most common example of positive perchlorate or iodide-perchlorate discharge tests is found in Hashimoto's thyroiditis (Morgans and Trotter, 1957; Murray and McGirr, 1960; Volpé et al., 1965). Dischargeable iodide is sometimes absent in this disorder but more often is associated with a partial organification defect. The literature on this defect in Hashimoto's thyroiditis is extensive and will not be reviewed here except to note that in mild cases it may require the iodide-perchlorate test to evoke a positive result. The great sensitivity of Hashimoto's thyroiditis toward iodide in inducing thyroid blockade probably describes the same phenomenon (viz. Wolff, 1969). Despite the fact that a positive perchlorate discharge test is often observed in Hashimoto's thyroiditis, the details of the defect are not understood. Because of the frequent presence of anti-TPO antibodies, some have assumed that the antibody may have damaged or inhibited the peroxidase in the residual functioning thyroid tissues. This has not, to our knowledge, been demonstrated. One could postulate that with slow cell damage, functions at the basolateral membrane might survive longer than the organification system, for example. Such dissociation has, in fact, been observed after irradiation of thyroid tissue (Barzelatto et al., 1962; Maloof et al., 1952), but information for thyroiditis is not available. It has also been suggested that Hashimoto thyroid tissue may iodinate thyroglobulin but not catalyze the coupling reaction to form T₃ and T₄ (Medeiros-Neto and Stanbury, 1994). This is, however, not readily compatible with the positive discharge tests. Women with previous postpartum thyroiditis may show persistent positive iodide-perchlorate discharge tests when they have gone through both the hyper and the hypothyroid phases, but not when they suffered only from the transient hyperthyroid phase (Creagh et al., 1994).

WOLFF

E. Ectopic Thyroid Tissue

Lingual thyroids may exhibit any level of thyroid function from normal to no function at all, and they may be malignant. It is thus not surprising that organification defects have been found. In 9 of 20 cases of lingual thyroid, positive perchlorate discharge tests were measured (Ferrini and Biassoni, 1966). Interestingly, in one of these with both in situ and ectopic tissue, only the lingual tissue showed a positive test. In 13 lingual or sublingual locations, the mean perchlorate discharge of ¹²³I was 43% (Hilditch and Jackson, 1985). Pertechnetate can be similarly discharged by perchlorate from lingual thyroids or ectopic tissue, as visualized by scintigraphy (Al-Jurayyan and El-Desouki, 1997; El-Desouki et al., 1995). As with some congenital thyroids, the defect may be transient. Finally, a positive discharge test was shown scintigraphically in a follicular and spindle cell carcinoma metastasis to the skull (Valenta, 1966) (see table 2; note that some items are classified under several headings; in addition, a great many references to discharge tests are left out for reasons of

Finally, a less well-known use for perchlorate, not directly linked to thyroid function but illustrating the use of lessons learned from iodide-concentrating tissues, generally, such as salivary glands, choroid plexus, gastric mucosa, etc., is its use in pertechnetate (99mTcO₄⁻) scintigraphy, for example, in the brain. Contaminating pertechnetate in organic technetium reagents tends to enter the parotid gland. Prevention of this by perchlorate improves the images (Ballinger et al., 1990). Because pertechnetate and perchlorate are both competitors for the Na/I⁻ symporter in all tissues where it is found, perchlorate may be used to identify certain choroid plexus papillomas. The pertechnetate that has been accumulated in these structures can be discharged by perchlorate, whereas in other tumors or in the rest of the brain, labeled technetium cannot be discharged (Cleto et al., 1992). Perchlorate is similarly useful in pertechnetate investigation of gastrointestinal bleeding (Hilditch et al., 1985).

IX. Summary

Perchlorate competitively blocks iodide from entering the thyroid by an effect on the Na⁺/I⁻ symporter thus preventing the further synthesis of thyroid hormone but has no effect on the iodination process itself. It is concentrated by thyroid tissue in a manner similar to iodide but is not significantly metabolized in the gland or peripherally. What is not settled is whether there are additional perchlorate effects on iodide transport. Perchlorate has a fast turnover in the body and requires frequent daily doses for therapy of thyrotoxicosis. Perchlorate appears to be substantially more effective against large iodide loads than the thionamides, and, with long-term iodide contamination, combined therapy of perchlorate (with ≤ 1 g/day) and thionamides is recommended for the more severe cases of thyrotoxicosis that may result from excess iodide or iodide-generating organic compounds, as for example with amiodarone. After ~ 30 days, the perchlorate dosage can be tapered or stopped, continuing with thionamides alone. This markedly increases its safe use. Despite serious side effects during its early use, lower dosages and shorter treatment periods appear to have prevented such reactions in its recent reintroduction, mostly for amiodarone-induced thyroid dysfunction. Perchlorate can also protect against inhibition of thyroid function and the resulting hypothyroidism caused by excess iodide, presumably by reducing the formation of an iodinated inhibitor. The reduction of the iodide pool by perchlorate thus has dual effects—reduction of excess hormone synthesis and hyperthyroidism, on the one hand, and reduction of thyroid inhibitor synthesis and hypothyroidism on the other. Perchlorate remains very useful also as a single dose application in tests measuring the discharge of radioiodide accumulated in the thyroid as a result of many different disruptions in the further metabolism of iodide in the thyroid gland.

Acknowledgments. I am most grateful to the following for careful criticisms of the manuscript: Dr. Jacob Robbins, NIH; Dr. John B. Stanbury, Boston; and Dr. Wilmar M. Wiersinga, Amsterdam.

Note: Since submission of this paper, an outstanding and extensive review on amiodarone and the thyroid has been published by W. M. Wiersinga (in Handbook of Experimental Pharmacology 128: 225-287, 1997), which the reader should consult for additional information. In addition, interesting findings regarding anion selectivities have been published for the symporter expressed in Xenopus oocytes (Eskandari, S., Loo, D. D. F., Dai, G., Levy, O., Wright, E. M., and Carrasco, N.: J Biol Chem 272:27230-27238, 1997). These selectivities differ from nearly all previously published material in that perchlorate, and the closely related perrhenate, ions are not transported, whereas accumulation in thyroid tissue has been repeatedly demonstrated. By contrast, anions weakly active in the thyroid, such as chlorate and nitrate, were well transported in the oocyte system. These discrepancies may be due to differences in the number of compartments; differences in the metabolism of some of the anions; the high anion concentrations used; or the requirement for an additional factor for accumulation. It will be instructive to resolve these differences.

REFERENCES

Aanderud S, Sundsfjord J and Aarbakke J (1984) Amiodarone inhibits conversion of T₄ to T₃ in isolated rat hepatocytes. Endocrinology 115:1605–1608.

Al-Jurayyan NAM and El-Desouki MI (1997) Transient iodine organification defect in infants with ectopic thyroid glands. Clin Nucl Med 22:13-16.

Alberto R, Cerdan A and Sanchez-Franco F (1987) Congenital hypothyroidism from complete iodide transport defect: Long-term evolution with iodide treatment. Postgrad Med J 63:1043-1047

Downloaded from

pharmrev

.aspetjournals.org

Ò

guest

9

June

5

20

12

- Alexander NM (1959) Iodide peroxidase in rat thyroid and salivary glands and its inhibition by antithyroid compounds. J Biol Chem 234:1530-1533.
- Alexander WD and Wolff J (1964) Cation requirements for iodide transport. Arch Biochem Biophys 106:525-526.
- Alexander WD and Wolff J (1966) Thyroidal iodide transport: Relation between transport, goitrogenic and antigoitrogenic properties of certain anions. *Endocrinology* 78:581-590.
- Allen EM (1993) Acute iodine ingestion increases intrathyroidal glutathione. J Endocrinol Invest 16:265–270.
- Anbar M, Guttmann S and Lewitus Z (1959) The mode of action of perchlorate ions on the iodine uptake of the thyroid gland. Int J Appl Radiat Isot 7:87–96.
- Andersen BF (1973) Iodide perchlorate discharge test in lithium-treated patients.

 Acta Endocrinol 73:35-42.
- Astwood EB (1943) The chemical nature of compounds which inhibit the function of the thyroid gland. J Pharmacol Exp Ther 78:79–89.
- Bagchi N and Fawcett DM (1973) Role of sodium ion in active transport of iodide by cultured thyroid cells. Biochim Biophys Acta 318:235–251.
- Bakker O, Van Beeren HC and Wiersinga WM (1994) Desethylamiodarone is a noncompetitive inhibition of the binding of thyroid hormone to the β_1 receptor protein. *Endocrinology* 134:1665–1670.
- Ballinger JR, Gulenchyn KY and Reid RH (1990) Radiopharmaceutical factors in the variable quality of [⁹⁹mtc]-Hm-Pao images of the brain. *J Nucl Med* **31:**118–122. Bartalena L, Brogioni S, Grasso L, Bogazzi F, Burelli AA and Martino E (1996)
- Bartalena L, Brogioni S, Grasso L, Bogazzi F, Burelli AA and Martino E (1996) Treatment of amiodarone-induced thyrotoxicosis, a difficult chalange: Results of a prospective study. *J Clin Endocrinol & Metab* 81:2930–2933.
- Barzelatto J, Murray IPCA and Stanbury JB (1962) Effects of gamma radiation of oxygen utilization, iodine metabolism and leucine incorporation by surviving sheep thyroid slices. *Endocrinology* **70:**328–332.
- Barzilai DA and Sheinfeld M (1966) Fatal complications following the use of potassium perchlorate in thyrotoxicosis. *Isr J Med Sci* 2:453–456.
- Baschieri L, Benedetti G, deLuca F and Negri M (1963) Evaluation and limitations of the perchlorate test in the study of thyroid function. *J Clin Endocrinol & Metab* 23:786–791.
- Bastomsky CH (1974) Thyroid iodide transport, in *Handbook of Physiology, Section* 7: Endocrinology Vol III Thyroid (Greer MA and Solomon DH, eds) pp 81–99, American Physiological Society, Washington, D. C. Batsakis JG, Nishiyama RH and Schmidt RW (1963) "Sporadic goiter syndrome": A
- Batsakis JG, Nishiyama RH and Schmidt RW (1963) "Sporadic goiter syndrome": A clinicopathologic analysis. Am J Clin Path 39:241–251.
- Baumann EJ and Metzger N (1949) Behavior of the thyroid toward elements of the seventh periodic group: Halogens and thiocyanates. *Proc Soc Exp Biol Med* **70:** 536–540.
- Baumann EJ, Searle NZ, Yalow AA, Siegel E and Seidlin SM (1956) Behavior of the thyroid toward elements of the seventh periodic group. Am J Physiol 185:71–76. Bax GM (1966) Typical and atypical cases of Pendred's Syndrome in one family. Acta
- Bax GM (1966) Typical and atypical cases of Pendred's Syndrome in one family. Acta Endocrinol 53:264–270.
 Beddows SA, Page SR, Taylor AH, McNerney R, Whitley GS, Johnstone AP and
- Nussey SN (1989) Cytotoxic effects of amiodarone and desethylamiodarone on human thyrocytes. *Biochem Pharmacol* **38**:4397–4403.

 Belshaw BE and Becker DV (1973) Necrosis of follicular cells and discharge of iodine
- induced by administering iodide to iodine-deficient dogs. J Clin Endocrinol & Metab 36:466-474.
- Blørstad O and Vogt JH (1962) Combined treatment of thyrotoxicosis with perchlorate and propylthiouracil. Acta Med Scand 171:283–288.
- Brennan MD, Erickson DZ, Carney JA and Bahn RS (1995) Nongoitrous amiodarone-associated thyrotoxicosis: Evidence of follicular disruption in vitro and in vivo. Thyroid 5:177–183.
- Broekhuysen J, Laruel R and Sion R (1969) Research on the benzofuran series xxxviii: Comparative study of transit and metabolism of amiodarone in different species of animals and humans. *Arch Int Pharmacodyn* 177:340–359.
- Brown-Grant K (1961) Extrathyroidal iodide concentrating mechanisms. *Physiol Rev* 41:189–213.
- Burger A, Dinichert D, Nicod P, Jenny M, Lemarchand-Béraud T and Vallotton MD (1976) Effect of amiodarone on serum T₃, rT₃, T₄ and Tsh. J Clin Invest **58:**255–259
- Capiello F, Boldorini R, Tosoni A, Praneo S, Bernasconi R and Raggi U (1995) Ultrastructural evidence of thryoid damage in amiodarone-induced thyrotoxicosis. J Endocrinol Invest 18:862–868.
- Carrasco N (1993) Iodide transport in the thyroid gland. Biochim Biophys Acta 1154:65-82.
- Childs DS Jr, Keating FR Jr, Rall JE, Williams MD and Power MH (1950) The effect of varying quantities of inorganic iodide (carrier) on the urinary excretion and thyroidal accumulation of radioiodine in exophthalmic goiter. J Clin Invest 29: 726-738.
- Chiovato L, Martino E, Tonacchera M, Chiovato L, Martino E, Tonochera M, Santini F, Lapi F, Mammoli C, Braverman LE and Pinchera A (1994) Studies on the in vitro cytotoxic effect of amiodarone. *Endocrinology* **134**:2277–2282.
- Chow SY and Woodbury DM (1970) Kinetics of distribution of radioactive perchlorate in rat and guinea pig thyroid glands. *J Endocrinol* 47:207–218.

 Chow SY, Chang LR and Yen MS (1969) A comparison between the uptakes of
- Chow SY, Chang LR and Yen MS (1969) A comparison between the uptakes of radioactive perchlorate and iodide by rat and guinea pig thyroid glands. J Endocrinol 45:1–8.
- Clark OH, Cavalieri RR, Moser C and Ingbar SH (1990) Iodine-induced hypothyroidism in patients after thyroid resection. Eur J Clin Invest 20:573–580.
- Cleto EM, Holmes RA, Singh A, Bierman R, Islam S and Hoffman TJ (1992) Radiographic and neuro-SPECT imaging in an immature third ventricle teratoma. J Nucl Med 33:435–437.
- Collins KDA and Washabaugh MW (1985) The Hofmeister effect and the behaviour of water at interfaces. *Q Rev Biophys* 18:323–422.
- Connel JMC (1981) Long-term use of perchlorate. Postgrad Med 57:516-517.
- Cook WHR and Hawe P (1960) Perchlorate and the treatment of thyrotoxicosis. Lancet 1:545.

- Couch RM, Dean HJ and Winter JSD (1985) Congenital hypothyroidism caused by defective iodide transport. J Pediatr 106:950–953.
- Creagh FM, Parkes AB, Lee A, Adams H, Hall R, Richards CJ and Lazarus JH (1994) The iodide-perchlorate discharge test in women with previous postpartum thyroiditis: Relation to sonographic appearance and thyroid function. Clin Endocrinol 40:765–768
- Crooks J and Wayne EJ (1960) A comparison of potassium perchlorate, methylthiouracil and carbimazole in the treatment of thyrotoxicosis. *Lancet* 1:401–404.
- Crooks J, Wayne EJ and Robb RA (1960) A clinical method of assessing the results of therapy in thyrotoxicosis. *Lancet* 1:397–401.
- Dai G, Levy O and Carrasco N (1996) Cloning and characterization of the thyroid iodide transporter. Nature 379:458-460.
- DeGroot LJ and Bühler U (1971) Effect of perchlorate and methimazole on iodine metabolism. Acta Endocrinol 68:696-706.
- Edwards DAW, Rowlands EN and Trotter WR (1954) The mechanism of the goitrogenic action of p-aminosalicylic acid. *Lancet* 267:1051–1052.
- Eichler O and Hackenthal E (1962) Über ausscheidung und stoffwechsel von perchlorate gemessen mit ³⁶ClO₄. Naunyn-Schmiedebergs Arch Exp Pathol Pharmakol 243:554-565.
- Eisenman G (1961) On the elementary atomic origin of equilibrium ionic specificity, in *Membrane Transport and Metabolism* (Kleinzeller A and Kotyk A, eds) pp 163–179, Academic Press, New York.
- Eisenman G (1962) Cation selective glass electrodes and their mode of operation. Biophys J Suppl. 2: 259–323.
- Ekholm R (1990) Biosynthesis of thyroid hormones. Int Rev Cytol 120:242-288.
- El-Desouki M, Al-Jurayyan N, Al-Nuaim A, Al-Herbish A, Abo-Bakr A, Al-Mazrou Y and Al-Swailem A (1995) Thyroid scintigraphy and perchlorate discharge test in the diagnosis of congenital hypothyroidism. Eur J Nucl Med 22:1005–1008.
- Enneker C, Kessler FJ and Krüskemper HL (1958) Der einfluss von T₃ und T₄ auf die perchlorat struma. *Acta Endocrinol* **29:**565–574.

 Ferrini O and Biassoni P (1966) Crintatiraidisma enzimonenico. *Folia Endocrinol*
- Ferrini O and Biassoni P (1966) Criptotiroidismo enzimopenico. Folia Endocrinol 19:313–333.
- Floyd JC Jr, Beierwaltes WH, Dodson VN and Carr EA (1960) Defective iodination of tyrosine: A cause of nodular goiter? J Clin Endocrinol & Metab 20:881–888. Fradkin JE and Wolff J (1983) Iodide-induced thyrotoxicosis. Medicine 62:1–20.
- Fragu P, Schlumberger M, Davy J-M, Slama M and Berdeaux A (1988) Effects of amiodarone therapy on thyroid iodine content as measured by X-ray fluoresence. J Clin Endocrinol & Metab 66:762–769.
- Franklin AL, Chaikoff IL and Lerner SR (1944) The influence of goitrogenic substances on the conversion in vitro of inorganic iodide to thyroxine and diiodotyrosine. *J Biol Chem* **153**:151–162.
- Franklyn JA, Davis JR, Gamage MD, Littler WA, Ramsden DB and Sheppard MC (1985) Amiodarone and thyroid hormone action. Clin Endocrinol 22:257–264.
- Fraser GR, Morgans ME and Trotter WR (1960) The syndrome of sporadic goiter and congenital deafness. Q J Med 29:279–295.
- Friis J (1987) The perchlorate discharge test with and without supplement of KI. J Endocrinol Invest 10:581–584.
- Fritzsche H, Benzer W, Furlan W, Hammerle D, Langsteger W and Weiss O (1993)
 Zur vorbeugung jodinduzierter hyperthyreosen nach koronarangiographie. Acta
 Med Austriaca 20:13–17.
- Furth ED, Carvalho M and Vianna B (1967) Familial goiter due to an organification defect in euthyroid siblings. J Clin Endocrinol & Metab 27:1137–1140.
- Gassner FX, Hopwood ML, Herrold EA and Plummer AJ (1950) An interpretation of the goitrogenic properties of certain antithyroid agents. J Clin Endocrinol & Metab 10:1485–1498.
- Gjemdal N (1963) Fatal aplastic anemia following use of potassium perchlorate in thyrotoxicosis. Acta Med Scand. 174:129-131.
- thyrotoxicosis. Acta Med Scand 174:129–131.
 Godley AF and Stanbury JB (1954) Preliminary experience in the treatment of hyperthyroidism with potassium perchlorate. J Clin Endocrinol & Metab 14:70–78.
- Goldberg RC and Wolff J (1954) Evaluation of the antithyroid activity of 5-iodo-2thiouracil. Endocrinology 54:181–195.
- Goldman SJ and Stanbury JB (1973) The metabolism of perchlorate in the rat.
 Endocrinology 92:1536-1538.
 Gray HW, Hooper LA and Greig WR (1973) An evaluation of the twenty minute
- perchlorate discharge test. J Clin Endocrinol & Metab 37:351–355.
 Haffajee CI, Love JC, Canada AT, Asdourian G and Alpert SS (1985) Clinical
- Haffajee CI, Love JC, Canada AT, Asdourian G and Alpert SS (1985) Clinical pharmacokinetics and efficacy of amiodarone. Circulation 67:1347–1355.
- Halmi NS (1961) Thyroidal iodide transport. Vitam Horm 19:133–163.
- Halmi NS, Granner DK, Müller G, Peters BH and Smith BD (1960) Effect of thiocyanate, stable iodide and perchlorate on the kinetics of radioiodide transport between thyroid gland and blood of rats. Endocrinology 67:332-336.
 Hariai KJ and Licato AA (1997) Effects of amiodarone on thyroid function. Ann
- Intern Med 126:63–73.

 Haust HL and Heagy FC (1980) Abnormal perchlorate-induced radioiodide loss from
- Haust HL and Heagy FC (1980) Abnormal perchlorate-induced radioiodide loss from the thyroid gland in porphyrinopathies. Int J Biochem 12:981–984.
- Hawthorne GC, Campbell NP, Geddes JS, Ferguson WR, Postlethwaite W and Sheridan B (1985) Amiodarone-induced hypothyroidism. Arch Intern Med 145: 1016–1019.
- Hilditch TE, Horton PW, McCruden DC, Young RE and Alexander WD (1982) Defects in intrathyroidal binding of iodine, and the perchlorate discharge test. Acta Endocrinol 100:237–244.
- Hilditch TE and Jackson HJ (1985) Quantitative ¹²³I-iodide scintigraphy and radiation dosimetry in infants with congenital hypothyroidism. *Eur J Nucl Med* 11:132–135
- Hilditch TE, Birnie GG, Sik MJ and Gillen G (1985) Use of perchlorate to block gastric uptake of free ⁹⁹mTc in the investigation of gastric bleeding. *Nucl Med Commun* **6:**701–706.
- Holt DW, Tucker GT, Jackson PR and Storey GCA (1983) Amiodarone pharmacokinetics. Am Heart J 106:840–847.
- Iff HW and Wilbrandt W (1963) Die abhängigkeit der jodakkumulation in schild-

104 WOLFF

drüsenschnitten von der ionalen zusammensetzung des inkubationsmediums und ihre beeinflussung durch herzglykoside. Biochim Biophys Acta 78:711-725.

Inada M, Nishikawa M and Kawai I (1983) Hypothyroidism associated with positive results of the perchlorate discharge test in elderly patients. Am J Med 74:1010-1015

- Ingbar SH and Freinkel N (1956) Concentration gradients for radioiodide in unblocked thyroid glands of rats: Effect of perchlorate. Endocrinology 58:95–103.
- Jolin T, Morreale de Escobar G and Escobar del Rey F (1968) 6-Propyl-2-thiouracil vs perchlorate-induced goiters. Endocrinology 83:620-625.
- Jolin T, Morreale de Escobar G and Escobar del Rey F (1970) Differential effects in the rat of thyroidectomy, propylthiouracil and other goitrogens on plasma insulin and thyroid weight. Endocrinology 87:99-110.
- Jolin T, Tarin MJ and Garcia MD (1974) Effect of adrenalectomy and corticosterone on thyroid weight of goitrogen-treated rats. Acta Endocrinol 75:734-747
- Jonekheer MH and Deconinck F (1982) X-Ray fluorescence determination of stable iodine in the thyroid gland. Acta Clin Belg 37:30-40, 92-102.
- Joseph U, Schmidt JA, Ehlenz K, Leppek R, Rothmund M, Wichert P and Joseph K (1994) Jodinduzierte hyperthyreose bei metastasierten schilddrüsen karzinom. Dtsch Med Wschr 119:1573-1578.
- Kahl S and Bobek S (1971) Antigoitrogenic properties of some anions in animals treated with propylthiouracil. Pol Endocrinol 22:420-428.
- Kasukabe T (1975) Deficient cytochrome b_5 reductase activity in nontoxic goiter with iodide organification defect. Metabolism~24:1103–1113.
- Kieffer J, Medeiros-Neto GA and Rueda R (1965) Perchlorate test in hyperthyroid patients treated with radioactive iodine. New Engl J Med 273:1326-1327.
- Koutras DA, Souvatzoglou A, Pandos PG, Papachristou DN, Piperingos GD and Sfontouris J (1978) Iodide organification defect in iodine deficiency endemic goiter. J Clin Endocrinol & Metab 47:610-614.
- Krevans JR, Asper SP and Rienhoff WF (1962) Fatal aplastic anemia following use of potassium perchlorate in thyrotoxicosis. J Am Med Assoc 181:162–164.
- Krüskemper HL (1960a) Nebenwirkungen thyreostatischer arzneimittel. Internist
- Krüskemper HL (1960b) Theoretische grundlagen und klinische ergebnisse der
- behandlung von hyperthyreosen mit perchlorat. Arzneim Forsch 10:13-17. Krüskemper HL (1962) Perchlorat als ursache von leukopenie, agranulozytose und
- aplastischer anämie. Deut Med Wchschr 87:1427-1431. Krüskemper HL and Kleinsorg H (1954) Antithyroideale wirkung von perchlorat.
- Naunyn-Schmiedebergs Arch Exp Pathol Pharmakol 223:469–480.
 Kugler HK (1969) Chlor -Das ClO₄ ¯ ion, in Gmelins Handbuch der Anorganischen
- Chemie, 8th ed, No 6, Ergänzungsband, Part B, Vol 2, pp 479-482, Verlag Chemie GMBH, Weinheim, Germany.
- Kutzim H, Mödder G and Buschsieweke U (1980) Iodine kinetics in facultative hyperthyroidism. J Mol Med 4:75-86.
- Lambert M, Unger J, Denayer P, Brohet C and Gangji D (1990) Amiodarone-induced thyrotoxicosis suggestive of thyroid damage. J Endocrinol Invest 13:527-530.
- Latini R, Tognoni G and Kates RE (1984) Clinical pharmacokinetics of amiodarone. Clin Pharmacokinet 9:136-156.
- Léger FA, Fragu P, Rougier P, Laurent MF, Tubiana M and Savoie JC (1983) Thyroid iodine content measured by x-ray fluorescence in amiodarone-induced thyrotoxicosis. J Nucl Med 24:582-585.
- Léger FA, Massin JP, Laurent MF, Vincens M, Auriol M, Helal OB, Chouette G and Savoie JC (1984) Iodine-induced thyrotoxicosis: Analysis of 85 consecutive cases. Eur J Clin Invest 14:449-455.
- Léger FA, Doumith R, Courpotin C, Helal OB, Davous N, Aurengo A and Savoie JC (1987) Complete iodide trapping defect - two cases of congenital hypothyroidism: Adaptation of thyroid to huge iodide supplementation. Eur J Clin Invest 17:249-
- Lever EC, Medeiros-Neto GA and DeGroot LJ (1983) Inherited disorders of thyroid metabolism. Endocr Rev 4:213-239.
- Lewitus Z, Guttmann S and Anbar M (1962) Effect of thyroid-stimulating hormone (TSH) on the accumulation of perchlorate and fluoroborate ions in the thyroid glands of rats. Endocrinology 70:295-297.
- Maayan ML and Rosenberg IN (1968) Effects of ethionine on thyroid function. Endocrinology 82:1223-1226.
- Mackenzie CG (1947) Differentiation of the antithyroid action of thiouracil, thiourea and Paba from sulfonamides by iodide administration, Endocrinology 40:137-153. Maloof F, Dobyns BM and Vickery AL (1952) The effects of various doses of radioactive iodine on the function and structure of the thyroid of the rat. Endocrinology
- **50:**612-638. Many MC, Mestdagh C, Van Den Hove M-F and Denef J-F (1992) In vitro study of acute toxic effects of high iodine doses in human thyroid follicles. Endocrinology **131:**621-630
- Martino E. Aghini-Lombardi F. Mariotti S. Bartalena L. Braverman L and Pinchera A (1987a) Amiodarone: A common source of iodine-induced thyrotoxicosis. Horm
- Martino E, Aghini-Lombardi F, Mariotti S, Bartalena L, Lenziardi M, Ceccarelli C, Bambini G, Safran M, Braverman LE and Pinchera A (1987b) Amiodarone iodineinduced hypothyroidism: Risk factors and follow-up in 28 cases. Clin Endocrinol **26:**227–239
- Martino E. Aghini-Lombardi F. Mariotti S. Lenziardi M. Baschieri L. Braverman LE and Pinchera A (1986a) Treatment of amiodarone-induced thyrotoxicosis by simultaneous administration of potassium perchlorate and methimazole. J Endocrinol Invest 9:201-207.
- Martino E, Bartalena L, Mariotti S, Anghini-Lombardi F, Ceccarelli C, Lippi F, Piga M, Loviselli A, Braverman L, Safran M and Pinchera A (1988) Radioactive iodine thyroid uptake in patients with amiodarone-induced thyroid dysfunction. Acta Endocrinol 119:167-173.
- Martino E, Mariotti S, Aghini-Lombardi F, Lenziardi M, Morabito S, Baschieri L Pinchera A, Braverman L and Safran M (1986b) Short term administration of potassium perchlorate restores euthyroidism in amiodarone iodine-induced hypothyroidism. J Clin Endocrinol & Metab 63:1233-1236.

- Martino E, Safran M, Aghini-Lombardi F, Rajatanavin R, Lenziardi M, Fay M, Pacchiarotti A, Aronin N, Macchia E, Haffajee C, Odoguardi L, Love J, Bigali A, Baschieri L, Pinchera A and Braverman L (1984) Environmental iodine intake and thyroid dysfunction during chronic amiodarone therapy. Ann Intern Med 101:28-34
- Mason JW (1987) Amiodarone. New Eng J Med 316:455-466.
- Massart C, Hody B, Condé D, Leclech G and Nicol M (1989) Effect of amiodarone and propranolol on the functional properties of human thyroid follicles in collagen gel. Mol Cell Endocrinol 62:113-117.
- Matsuzaki S and Suzuki M (1981) Paradoxical effect of perchlorate on thyroid weight, glucose-6-phosphate dehydrogenase and polyamines in methylthiouraciltreated rats. Acta Endocrinol 97:491-495.
- Mazonson PD, Williams ML, Cantley LK, Dalldorf FG and Foster DR (1984) Myxedema coma during long-term amiodarone therapy. Am J Med 77:751–754.
- McGirr EM, Hutchinson JH and Clement WE (1959) Sporadic goitre due to dyshormonogenesis: Impaired utilization of trapped iodide. Scott Med J 4:107-114.
- Medeiros-Neto G and Stanbury JB (1994) Inherited Disorders of the Thyroid System. CRC Press, Boca Raton.
- Melmed S, Nademanee K, Reed AW, Hendrickson J, Singh BN and Hershman JM (1981) Hyperthyroxinemia with bradycardia and normal Tsh secretion following chronic amiodarone administration. J Clin Endocrinol & Metab 83:997-1001.
- Milne K and Greer MA (1962) Comparison of the effects of propylthiogracil and sulfadiazine on thyroidal biosynthesis and the manner in which they are influenced by supplemental iodide. Endocrinology 71:580-587.
- Morgans ME and Trotter WR (1957) Defective organic binding of iodine by the thyroid in Hashimoto's thyroiditis. Lancet 272:553-555.
- Morgans ME and Trotter WR (1958) Association of congenital deafness with goitre. The nature of the thyroid defect. Lancet 1:607-609.
- Murray Ipc and McGirr EM (1960) Radioactive iodine studies in the diagnosis of Hashimoto's thyroiditis. Br Med J 1:838–841.
- Nademanee K, Piwonka RW, Singh BN and Hershman JM (1989) Amiodarone and

Downloaded

pharmrev

.aspetjournals.org

ō

g

June

5

20

- thyroid function. Prog Cardiovasc Dis 31:427-437. Nakamura Y, Kotani T and Ohtaki S (1990) Transcellular iodide transport and
- iodination on the apical plasma membrane by monolayer porcine thyroid cells cultured on collagen-coated filters. $J\ Endocrinol\ 126:275-281.$
- Nakamura Y, Ohtaki S and Yamazaki I (1988) Molecular mechanism for iodide transport by thyroid plasma luminal vesicles. J Biochem 104:544-549.
- Newnham HH, Topliss DJ, Legrand BA, Chosich N, Harper RW and Stockigt JR (1988) Amiodarone-induced hyperthyroidism: Assessment of the predictive value of biochemical testing and response to combined therapy with propylthiouracil and perchlorate. Aust NZ J Med 18:37-44.
- Nilsson M, Björkman U, Ekholm R and Ericson LE (1990) Iodide transport in primary cultured thyroid follicle cells. Eur J Cell Biol 52:270-281.
- Nilsson M and Ericson LE (1994) Effect of epidermal growth factor on basolateral iodide uptake and apical iodide permeability in filter-cultured thyroid epithelium. Endocrinology 135:1428-1436.
- Nose O, Harada T, Miyai K, Hata N, Ogawa M, Maki I, Kanaya S, Kimwia S, Shimizu K and Yabuuchi H (1985) Transient neonatal hypothyroidism probably related to immaturity of thyroidal iodine organification. J Pediatr 108:573-576.
- O'Neill B, Magnolato D and Semenza G (1987) The electrogenic Na⁺-dependent I transport system on plasma membrane vesicles from thyroid gland. Biochim Biophys Acta 896:263-274.
- Ortiz-Caro J, Pastor RM and Jolin T (1983) Effects of KClO4 in propylthiouraciltreated rats. Acta Endocrinol 103:81-87.
- Pereira A, Brackman J-C, Dumont JE and Boeynaems J-M (1990) Identification of a major iodolipid from horse thyroid gland as 2-iodohexadecanal. J Biol Chem **265:**17018-17025
- Pitt-Rivers R. Galton VA and Halmi NS (1958) Nature of radioiodine not dischargeable with perchlorate in the thyroid glands of thiouracil-treated rats. Endocrinology 63:699-703.
- Postel S (1957) Placental transfer of perchlorate and triiodothyronine in the guinea pig. Endocrinology 60:53-66.
- Raben MS (1949) The paradoxical effects of thiocyanate and of thyrotropin on the organic binding of iodine by the thyroid in the presence of large amounts of iodide. Endocrinology 45:296-304.
- Rao RH, McCready VR and Spathis CS (1986) Iodine kinetic studies during amiodarone treatment. J Clin Endocrinol & Metab 62:563-568.
- Reichenberg D (1966) Ion exchange selectivity, in Ion Exchange (Marinsky JA, ed), Vol I, pp 227-276, Marcel Dekker, New York.
- Reichert LJM and deRooy HAM (1989) Treatment of amiodarone-induced hyperthyroidism with potassium perchlorate and methimazole during amiodarone treatment. Br Med J 298:1547-1548
- Rocmans PA, Penel JC, Cantraine FR and Dumont JE (1977) Kinetic analysis of iodide transport in dog thyroid slices: Perchlorate-induced discharge. Am J Physiol
- Røkke KE and Vogt JH (1968) Combination of potassium perchlorate and propylthiouracil in the treatment of thyrotoxicosis. Acta Endocrinol 57:565-577
- Roti E, Minelli R, Gardini E, Bianconi L, Gavaruzzi G, Ugolotti G, Neri TM and Braverman LE (1992) Iodine-induced subclinical hypothyroidism in euthyroid subjects with a previous episode of amiodarone-induced thyrotoxicosis. J Clin Endocrinol & Metab **75:**1273–1277.
- Roti E, Minelli R, Gardini E, Bianconi L, Salvi M, Gavaruzzi G, Ugolotti G and Braverman LE (1994) The iodine perchlorate discharge test before and after one year of methimazole treatment of hyperthyroid Graves' Disease. J Clin Endocrinol & Metab 78:795-799.
- Rousset B, Orgiazzi J and Mornex R (1977) Perchlorate ion enhances mouse thyroid responsiveness to thyrotropin, human chorionic gonadotropin and long acting thyroid stimulator. Endocrinology 100:1628-1635.
- Saito K, Yamamoto K, Nagayama I, Uemura J and Takeshi K (1989) Effect of internally loaded iodide, thiocyanate and perchlorate. J Biochem 105:790-793.
- Saito K, Yamamoto K, Takai T and Yoshida S (1982) The sodium-dependent iodide

Downloaded from

pharmrev

.aspetjournals.org

è

guest

9

June

5

2012

- transport by phospholipid vesicles reconstituted with the thyroid plasma membrane. J Biochem 92:2001-2007.
- Scranton JR and Halmi NS (1965) Thyroidal iodide accumulation and loss in vitro. Endocrinology 76:441-453.
- Shukla R, Jowett NI, Thompson DR and Pohl JEF (1991) Side effects with amiodarone therapy. Postgrad Med J 70:492-498.
- Singh BM and Nademanee K (1983) Amiodarone and thyroid function: Clinical implications during antiarrhythmic therapy. Am Heart J 106:857-869.
- Smanik PA, Liu Q, Furmiger TL, Ryu K, Xing S, Mazzaferri EL and Jhiang SM (1996) Cloning the human sodium iodide symporter. Biochem Biophys Res Commun 226:339-345.
- Smyrk TC, Goellner JR, Brennan MD and Carney JA (1987) Pathology of the thyroid
- in amiodarone-associated thyrotoxicosis. *Am J Surg Path* 11:197–204. Stanbury JB and Wyngaarden JB (1952) Effect of perchlorate on the human thyroid gland. Metabolism 1:533-539.
- Stanley MM and Astwood EB (1948) The accumulation of radioactive iodide by the thyroid gland in normal and thyrotoxic subjects and the effect of thiocyanate on its discharge. Endocrinology 42:107-123.
- Stanley MM (1949) The direct estimation of the rate of thyroid hormone formation in man: The effect of the iodide ion on thyroid iodine utilization. J Clin Endocrinol & Metah 9:941-954
- Stanley MM and Astwood EB (1949) The response of the thyroid gland in normal human subjects to the administration of thyrotropin, as shown by studies with I¹³¹. Endocrinology 44:49-60.
- Stewart RDH and Murray IPC (1966) An evaluation of the perchlorate discharge test. J Clin Endocrinol & Metab 26:1050-1058.
- Stewart RDH and Murray IPC (1967) Effect of small doses of carrier iodide upon the organic binding of radioactive iodine by the human thyroid gland. J Clin Endocrinol & Metab 27:500-508.
- Surks MI (1967) Determination of iodide clearance and exit rate constants in incubated thyroid lobes. Endocrinology 80:1020-1027.
- Takeuchi K, Suzuki H, Horiuchi Y and Mashimo K (1970) Significance of iodideperchlorate discharge test for detection of iodine organification defect of the thyroid. J Clin Endocrinol & Metab 31:144-146.
- Taurog A (1996) Hormone synthesis and secretion, in Wemer and Ingbar's The Thyroid, (Braverman LE and Utiger RD, eds), 7th ed, pp 47-81, Lippincott-Raven, Philadelphia.
- Trip MD, Düren DR and Wiersinga WM (1994) Two cases of amiodarone-associated thyrotoxicosis successfully treated with a short course of antithyroid drugs while amiodarone was continued. Br Heart J 72:266-268.
- Trip MD, Wiersinga W and Plomp TA (1991) Incidence, predictability and pathogenesis of amiodarone-induced thyrotoxicosis and hypothyroidism, Am J Med 91:507-511
- Trotter WR (1960) The association of deafness with thyroid dysfunction. Br Med Bull
- Trotter WR (1962) The relative toxicity of antithyroid drugs. J New Drugs 2:333-343. Tyler DD, Gonze J, Lamy F and Dumont JE (1968) Influence of mitochondrial inhibitors on the respiration and energy-dependent uptake of iodide by thyroid slices. Biochem J 106:123-133.
- Valenta L (1966) Metastatic thyroid carcinoma in man concentrating iodine without organification. J Clin Endocrinol & Metab 26:1317-1324.
- Van Beeren HC, Bakker OLA and Wiersinga WM (1995) Desethylamiodarone is a competitive inhibitoof binding of T3 to the a1 receptor. M ol Cell Endocrinol
- Van Dam Ewcm, Prummel MF, Wiersinga WM and Nikkels RE (1993) Treatment of amiodarone-induced hypothyroidism with potassium perchlorate. Net J Med 42:
- Vanderlaan JE and Vanderlaan WP (1947) The iodide concentrating mechanism of the rat thyroid and its inhibition by thiocyanate. Endocrinology 40:403-416.
- Vilijn F and Carrasco N (1989) Expression of the thyroid sodium/iodide symporter in Xenopus laevis oocytes. J Biol Chem 264:11901-11903.
- Volpé R, Row VV, Webster BR, Johnston MAW and Ezrin C (1965) Studies of iodine metabolism in Hashimoto's thyroiditis. J Clin Endocrinol & Metab 25:593-601.

- Vulsma T, Rammeloo JA, Gons MH and deVijlder JJM (1991) The role of serum thyroglobulin concentration, and thyroid ultrasound imaging in the detection of iodide transport defects in infants. Acta Endocrinol 124:405-410.
- Wémeau JL, Bauters C, Gérard Y, Deligne A, Coviaux R, Lion G, Steinling M and Decoulx M (1991) Le perchlorate de potassium dans L'exploration et le traitemant des maladies thyroidiennes. Rev Fr Endocrinol Clin Nutr Metab 32:473-482.
- Wenzel KW and Lente JR (1984) Similar effects of thionamide drugs and P2 on Tsh in Graves' disease: Evidence against an immunosuppressive action of thionamide drugs J Clin Endocrinol & Metab 58:62-69
- Wiersinga WM, Touber JL, Trip MD and van Royen EA (1986) Uninhibited thyroidal uptake of radioiodine despite iodide excess in amiodarone-induced hypothyroidism. J Clin Endocrinol & Metab 63:485-491.
- Williams JA (1966) Effect of external K+ concentration on transmembrane potentials of rabbit thyroid cells. Am J Physiol 211:1171-1174.
- Wolff J (1964) Transport of iodide and other anions in the thyroid gland. Physiol Rev 44:45-90.
- Wolff J (1969) Iodide goiter and the pharmacologic effects of excess iodide. Am J Med 47:101-124
- Wolff J (1972) Iodide concentrating mechanism, in Methods in Investigative and Diagnostic Endocrinology: The Thyroid and Biogenic Amines (Rall JE and Kopin IJ, eds), Vol I, pp 115–140, North Holland Publishing, Amsterdam.
- Wolff J (1983) Congenital goiter with defective iodide transport. Endocr Rev 4:240-254
- Wolff J (1989) Excess iodide inhibits the thyroid by multiple mechanisms, in Control of the Thyroid Gland (Ekholm R Kohn L and Wollman SH, eds) pp 211-244, Plenum, New York.
- Wolff J (1992) Iodure: Le systeme de transport actif, in La Thyroide (Leclere J, Orgiazzi JU, Rousset BU, Schlienger J-L and Wémeau J-L, eds) pp 17-24, Expansion Scientique Française, Paris.
- Wolff J (1996) Iodide prophylaxis for reactor accidents, in Proceedings of the Nagasaki Symposium 1995, (Nagataki S and Yaurashita S, eds) pp 227-237, Elsevier, Amsterdam.
- Wolff J and Halmi NS (1963) Thyroidal iodide transport: The role of Na+-K+activated, ouabain-sensitive adenosinetriphosphatase activity. J Biol Chem 238:
- Wolff J and Maurey JR (1958) Cardiac glycosides and thyroidal iodide transport. Nature 182:957.
- Wolff J and Maurey JR (1962) Thyroidal iodide transport: Comparison of iodide with anions of periodic group VII A. Biochim Biophys Acta 57:422-426.
- Wolff J and Maurey JR (1963) Thyroidal iodide transport: The role of ion size. Biochim Biophys Acta 69:58-67.
- Wolff J, Robbins J and Rall JE (1959) Iodide trapping without organification in a transplantable rat thyroid tumor. Endocrinology 64:1-11.
- Wollman SH (1956) Nature of the inhibition by thiocyanate of the iodide concentrating mechanism of the thyroid gland. Am J Physiol 186:453-459.
- Wollman SH (1995) Thyroid radioiodide transport: Models, rate limiting steps and relation to formation of iodoprotein. Eur J Ĉell Biol $\mathbf{66:}217-225.$
- Woodbury DM and Woodbury JW (1963) Correlation of micro-electrode potential recordings with histology of rat and guinea pig thyroid glands. J Physiol 169:553-
- Wright EM and Diamond JM (1977) Anion selectivity in biological systems. Physiol Rev 57:109-156.
- Wyngaarden JB, Stanbury JB and Rapp B (1953) The effects of iodide, perchlorate, thiocyanate and nitrate administration upon the iodide concentrating mechanism of the rat thyroid. Endocrinology 52:568-574.
- Wyngaarden JB, Wright BM and Ways P (1952) The effect of certain anions upon the accumulation and retention of iodide by the thyroid gland. Endocrinology 50:537-
- Yamada T (1967) Effects of perchlorate and other anions on thyroxine metabolism in the rat. Endocrinology 81:1285-1290.
- Yamada T and Jones AE (1968) Effect of thiocyanate, perchlorate and other anions on plasma protein-thyroid hormone interaction in vitro. Endocrinology 82:47-53.